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How Good Is Your Lab?

By CRAIG PASKVAN

When you submit samples to your lab, how good are the results? Do you know if the samples you are submitting are going to be accurate? Can you count on making recommendations based on the results? What guidelines are there out there that you as a superintendent can use to assure yourself that you are dealing with a quality lab?

If they are in the upper 90%, you know you are dealing with a very good lab. If they are not, shop around and locate one that is a member of that group. Ask fellow superintendents where they have their work done. After all, how much is riding on those results?

(Editor's Note: Craig Paskvan is an independent soil consultant.)

The feeling I get from talking to different superintendents is that all labs are just about the same. Certainly in this day and age of automated equipment and robotic pH systems it makes repeated testing very easy to do. But the fact of the matter is that it takes good quality technicians and a strict quality control to produce accurate data on a routine basis. There are factory labs out there that have the best in modern equipment, perform high volumes of soil tests, yet fail to produce accurate results. All at a very low price. So can we assume that these labs are sacrificing speed for accuracy?? After all what is their goal? To provide the cheapest test possible, or produce the most accurate tests, ones on which you can depend?

One of the ways to determine if the lab you are using is a good quality lab is to determine if your lab belongs to the North American Proficiency Testing Program. There are about 152 plus labs that participate in this program each year. Samples are submitted to the labs for analysis which is done quarterly. Samples are flagged when the median average deviations become too high. Last year 13%, or 20 out of 152 labs, fell into the 90% passing category for accuracy. In other words, 20 out of 152 plus labs produced data that had 90% of the results not flagged. These labs are doing a very good job. The next level had 84 labs, or 55% of the 152 labs, fall into the 80 to 90% passed criteria. These are okay, but have considerable problems they need to address. And then you have the bottom 32% of the labs, or 49 labs, that fall below 70% passing. These labs have numerous concerns concerning their accuracy. It is surprising that state and government labs fall into this category, not just the private labs. These same problems are found in other labs around the world.

So what should you do if you are submitting samples to a lab which you make fertility recommendations on? Call the lab and verify that #1, they are a member of the North American Proficiency Testing program, and #2, find out where they rank.





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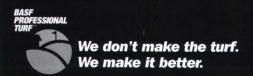


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BASF

History of Simplot Partners-

(Continued from Page 6)

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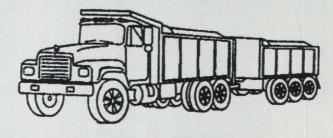
The company sells its turf and horticulture products worldwide through its own network as well as a large group of independent distributors. The Company's own distribution network, known as Simplot Partners, has 20 locations throughout the Untied States.. The Roseville, MN facility has been in operation since 2000 and currently serves the entire state of Minnesota and western Wisconsin. Besides selling proprietary products marketed by Jacklin Seed and the Best brand fertilizers, Simplot Partners enjoys strong positions with many major chemical companies including Syngenta, Dow AgroScience and PBI Gordon. Simplot Partners also carries its own line of specialty products including surfactants, liquid and water soluble fertilizers and a wide range of other nutrients and adjuvants. In the Spring of 2003, SP will be rolling out its own line of natural organic fertilizers as well.

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APRIL 2003 HOLE NOTES 19

Why Has It Been So Hard to Control Anthracnose These Past Few Years?

By Dr. Geunhwa Jung and Dr. Tae-hyun Chang Department of Plant Pathology, University of Wisconsin

Had problems controlling anthracnosedisease the past several summers? Well, there might be one possible explanation, Dr. Tae hyun Chang, who joined my lab in December 2001, carried out a field experiment this past summer to test fungicide efficacy for controlling anthracnose at the Blackhawk Country Club in Madison, WI. Dr. Chang came from South Korea and had played an important role in turf research by developing a pink snow mold inoculation method, assisting graduate students with experiments, analyzing field research data collected over the past years, and preparing a refereed publication. As one of the outcomes, he attended the annual national meeting of American Phytopathological Science (APS) last summer and gave an excellent poster presentation titled "Efficacy Fungicides on the control of Typhula Snow Molds in turfgrass". The manuscript is now being prepared for publication. His work with fungicides for anthtacnose will hopefully lead to equally beneficial results.

Recently anthracnose basal rot, caused by Colletotrichum graminicola, has become an important disease in Wisconsin and I have been repeatedly stating that the efficacy of some controlling fungicides has been reduced. Our lab had the opportunity of running an experiment with the cooperation of the superintendent at the Blackhawk Country Club. The objective of the study was to evaluate four systemic fungicides and one contact fungicide for their efficacy of controlling the disease. Hopefully, we will not only find the most efficacious fingicides, but also gain biological insight as to why the reduced efficacy in some fungicides was reported.

Materials and Methods

Fungicides labeled for controlling the anthracnose disease which were selected for this study are as follows: four systemic fungicides (Propiconazole, Triadimefon, Azoyxystrobin, and Thiophanatemethyl) one contact fungicide (Chlorothalonil), and one control (no chemical). the experimental plot was set up on fairway turf where more than 60% of the turf is estimated to be Poa annua species and the rest is creeping bentgrass. Over the years,

high disease pressure has been consistently observed at that location. Experimental design was a split plot with three apploication rates (low and high label rates anda mixture of systemic and contact fungicide with the respective low rate) as whole-plot treatments ina randomized complete block design. The fungicides were applied to subplots (Table 1). Individual plot size was 3 ft x 7 ft. Preventive chemical applications (14 day interval) were made on June 16, June 20, July 4 and July 18, 2002. Liquid treatments were applied witha CO2-powered boom sprayer using XR Teejet 8005 VS nozzles at 30 psi in water equivalent to 2 gal. per M sq. ft.

Disease ratings (percentage of infected area) of the plot were visually recoreded on August 2 and 17, 2002. The first disease symptom was noticed around the last week of July. in addition, the total percentage of Poa populations per plot was visually estimated twice. Since the anthracnose occurred only in Poa annua species, the percentage of the infected areas of Poa was recalculated by estimating the percentage of the entrie plot that was infected and then dividing that amount by the percentage of Poa present in the plot. The final data analysis using the recaclucated damage percentage wsa carried out and presented in (Table 1.)

Results

Tank mixing systemic and contact fungicides using their respective low label rate, had significantly higher efficacy for the anthracnose controle than the systemic fungicides alone regardless of the label rates (Table 1). However, the contact fungcide performed as well as the mixtures. More research is required for further confirmation of the results found in this (Continued on Page 21)

