Chapter 6.

A simple method for predicting cation exchange capacity, with special respect to sand rootzones

#### Abstract

The cation exchange capacity (CEC) of soil samples submitted to commercial soil testing laboratories is often estimated by summation of the nonacid cations (CEC<sub>sum</sub>) extracted in a routine soil nutrient analysis. These CEC estimates can predict measured CEC in non-calcareous soil samples but are inappropriate for use in calcareous soils and have not been evaluated in anthropogenic sand rootzones of turfgrass sites. In a dataset of 37 sand and 17 soil samples varying in pH, calcium carbonate equivalent (CCE), and organic matter (OM) content, we evaluated the ability of the summation method to predict measured  $CEC_{CE}$  (by compulsive exchange) using 5 extraction methods: 1 M NH<sub>4</sub>OAC, Mehlich 3, Morgan, 0.01 M SrCl<sub>2</sub>, and 1:5 H<sub>2</sub>O. The CEC<sub>sum</sub> estimates based on 1 M NH<sub>4</sub>OAC, Mehlich 3, and Morgan extractions were inaccurate for the calcareous samples and overestimated  $CEC_{CE}$  in sands with  $CCE < 10 \text{ g kg}^{-1}$ . The CEC<sub>sum</sub> based on the 1:5  $H_2O$  extraction underestimated CEC<sub>CE</sub> in all soil types. Estimates based on the 0.01 M SrCl<sub>2</sub> extraction were accurate in all sands and in soils with  $CEC_{CE} < 60 \text{ mmol}_{c} \text{ kg}^{-1}$ . In soils with  $CEC_{CE} > 60 \text{ mmol}_{c} \text{ kg}^{-1}$ , two consecutive extractions with 0.01 M SrCl<sub>2</sub> and summation of nonacid cations extracted provided an accurate estimate of CEC<sub>CE</sub>.

## Introduction

Most commercial soil testing laboratories estimate CEC from the quantity of nonacid cations extracted in an agronomic soil test (Ross, 1995; Sumner and Miller, 1996). Exchangeable acidity is also included in these estimates if the laboratory concludes that acidity is appreciable; in general, the exchangeable acidity can be considered negligible if soil pH in water is > 5.5.

In anthropogenic sand rootzones, extracting solutions such as Mehlich 3, Morgan, or 1 *M* NH<sub>4</sub>OAc are commonly used for extraction of nonacid cations, but the CEC estimates derived from nonacid cations extracted in these sand rootzones are of unknown accuracy. These sand rootzones are usually established with a minimum sand content of 920 g kg<sup>-1</sup> and maximum silt and clay content of 50 g kg<sup>-1</sup> and 30 g kg<sup>-1</sup>, respectively (USGA, 2004). Common extracting solutions such as Mehlich 3, Morgan, and 1 *M* NH<sub>4</sub>OAc have higher ionic strength (*I*) and a different pH than are found in soil solution, and these differences may influence the quantity of nonacid cations extracted from the organic matter which provides the majority of cation exchange sites in sand rootzones. Because sand rootzones are inherently low in exchangeable nonacid cations, any error in the measurement of exchangeable nonacid cations could have a relatively large influence on the estimated CEC. Furthermore, calcareous sands are geographically ubiquitous (Christians, 1990), but the usual method of estimating CEC based on the nonacid cations extracted with an agronomic extractant is clearly inappropriate for calcareous materials (Sumner and Miller, 1996).

There are numerous methods which make a more direct measurement of CEC (Bache, 1976; Gillman and Sumpter, 1986; Gillman et al., 1983; Matsue and Wada, 1985; Sumner and Miller, 1996), but they suffer from a common drawback that precludes their regular use: most methods are very time-consuming. For example, the BaCl<sub>2</sub> compulsive exchange method (CEC<sub>CE</sub>) of Gillman and Sumpter (1986), which determines the exchange of Mg<sup>2+</sup> for Ba<sup>2+</sup> at a known solution pH and *I*, requires multiple rinsing and equilibration steps that extend the procedure to 24 hours. Avila-Segura (1999) and Sumner et al. (1994) have suggested simplifications to the CEC<sub>CE</sub> method, but the method is in infrequent use at commercial soil testing laboratories, despite its accuracy, due to the length and complexity of the procedure.

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Matsue and Wada (1985) found that specific adsorption of  $SO_4^{2-}$  on andisols resulted in overestimation of  $CEC_{CE}$ , and recommended an alternative method involving sample equilibration with 0.01 *M* SrCl<sub>2</sub> in five consecutive centrifuge washings, after which adsorbed  $Sr^{2+}$  is removed by 0.5 *M* HCl and the removed  $Sr^{2+}$ measured as the CEC. This method requires 48 hours of equilibration and exchange, and therefore has not been widely used in commercial laboratories.

A single extraction with 0.01 M SrCl<sub>2</sub> has been used to measure exchangeable K (Woods et al., 2005a) and Ca and Mg (Woods et al., 2005b) of a calcareous sand rootzone. This rapid extraction adjusts to soil pH and has an I similar to that of soil solution.

For 37 sand and 17 soil samples, we compared the  $CEC_{CE}$  with CEC estimates derived from soil OM and pH and with CEC estimates calculated from summation of nonacid cations extracted by different extracting solutions: 1 *M* NH<sub>4</sub>OAC, Mehlich 3, Morgan, 0.01 *M* SrCl<sub>2</sub>, and 1:5 H<sub>2</sub>O. Our objective was to determine which of the CEC estimates could accurately predict CEC<sub>CE</sub> across a range of calcareous and noncalcareous sands and soils.

### **Materials and Methods**

Fifty-four sand (Table 6.1) and soil (Table 6.2) samples were obtained. Sands represented a range of pH, CCE, and OM typical of many turfgrass rootzones. Each sand sample was collected to a depth of 10 cm. The soils represented a range of pH, CCE, and OM; additionally, soil samples represented 5 of the 6 soil management groups (Bergstrom et al., 1987) in New York. Each soil sample was collected to a depth of 20 cm.

Sample Description	Location	рН	AI	OM	CCE	$CEC_{CE}$	
		(H <sub>2</sub> O)					
			mmol <sub>c</sub> kg⁻¹	g kg⁻¹	g kg⁻¹	mmol <sub>c</sub> kg <sup>-1</sup>	
Sand - golf green	Philippines	4.4	11	17	3	16	
Sand - golf green	Philippines	4.6	14	20	5	34	
Sand - golf fairway	New York	5.0	4	7	2	6	
Sand - golf green	Thailand	5.5	1	20	4	23	
Sand - golf green	Japan	5.7	1	17	5	11	
Sand - golf green	Japan	6.1		14	4	19	
Sand - golf fairway	Oregon	6.2		18	3	10	
Sand - golf green	New York	6.2		41	6	59	
Sand - golf green	Japan	6.4		20	7	7	
Sand - golf green	New York	6.5		37	7	45	
Sand - quarry	Unknown	6.5		0	2	0	
Sand - quarry	Unknown	6.6		4	2	0	
Sand - golf green	Oregon	6.7		14	6	30	
Sand - golf green	Oregon	6.8		10	3	19	
Sand - golf fairway	China	6.9		4	2	8	
Sand - golf green	Georgia	6.9		13	2	10	
Sand - golf tee	Thailand	7.0		11	4	27	
Sand - golf green	Japan	7.0		23	8	33	
Sand - golf green	Oregon	7.0		24	3	37	
Sand - golf green	Oregon	7.0		25	4	41	

Table 6.1. List of sands analyzed with associated data including pH, calcium carbonate equivalent (CCE), cation exchange capacity by compulsive exchange ( $CEC_{CE}$ ), and organic matter (OM).

Table	6.1	(Continued)	

Sand - golf green	California	7.2	15	7	40
Sand - golf green	New York	7.2	52	13	101
Sand - quarry	Unknown	7.3	2	2	0
Sand - golf green	Thailand	7.3	16	5	27
Sand - golf green	China	7.4	3	3	8
Sand - golf green	Spain	7.4	30	206	55
Sand - golf green	California	7.5	9	9	26
Sand - golf green	Japan	7.7	16	12	18
Sand - golf green	California	7.8	9	4	27
Sand - golf green	Japan	7.9	13	8	20
Sand - golf green	New York	8.2	8	241	14
Sand - golf green	New York	8.3	10	249	13
Sand - golf green	New York	8.3	11	233	13
Sand - golf rough	Hawaii	8.4	12	438	30
Sand - golf green	Arizona	8.5	5	18	30
Sand - golf green	Hawaii	8.8	6	740	9
Sand - quarry	New York	9.0	3	91	6

Soil Description	Location	рН	AI	OM	CCE	CEC <sub>CE</sub>
		(H <sub>2</sub> O)				
			mmol <sub>c</sub>	g kg⁻¹	g kg⁻¹	mmol <sub>c</sub> kg⁻¹
			kg⁻¹			
Fluventic Dystrudepts	New York	5.4	2	32	5	52
Typic Fragiudepts	New York	5.7	1	61	6	94
Typic Dystrudepts	New York	5.8	1	37	5	72
Typic Eutrudepts	New York	6.1		38	6	74
Typic Dystrudepts	New York	6.2		17	3	34
Fluventic Dystrudepts	New York	6.3		29	7	89
Aeric Endoaquepts	New York	6.4		37	6	71
Aeric Endoaqualfs	New York	6.7		47	8	191
Hudson clay loam	New York	6.7		53	7	79
Glossic Hapludalfs	New York	6.9		20	6	77
Glossic Hapludalfs	New York	7.0		36	10	109
Glossic Hapludalfs	New York	7.1		66	12	166
Glossic Hapludalfs	New York	7.1		106	19	172
Lamellic Hapludalfs	New York	7.2		28	20	108
Glossic Hapludalfs	New York	7.7		18	17	69

Table 6.2. List of soils analyzed with associated data including pH, calcium carbonate equivalent (CCE), cation exchange capacity by compulsive exchange (CEC<sub>CE</sub>), and organic matter (OM).

Table 6.2 (continued)

Aquic Udorthents	New York	7.7	40	10	131
Anthrosol	China	7.9	21	54	71

The CEC<sub>CE</sub> was measured using the modified compulsive exchange method of Gillman and Sumpter (1986). Nonacid cations were extracted from each sample with 1 M NH<sub>4</sub>OAc (Brown and Warncke, 1988), Mehlich 3 (Wolf and Beegle, 1995), and Morgan (Morgan, 1941). Water-soluble nonacid cations were extracted by 1:5 H<sub>2</sub>O (Soil and Plant Analysis Council, 1999).

Hundredth-molar SrCl<sub>2</sub> was substituted for 0.01 *M* CaCl<sub>2</sub> in the extraction method of Houba et al. (2000) to allow for direct measurement of Ca in the extract. The shaking time with 0.01 *M* SrCl<sub>2</sub> was reduced from 120 minutes to 5 minutes, and the soil:solution ratio was maintained at 1:10, but only 2 g soil and 20 mL 0.01 *M* SrCl<sub>2</sub> were used, rather than 10 g soil and 100 mL solution as described by Houba et al. (2000). This simplification of the 0.01 *M* SrCl<sub>2</sub> extraction was tested on 31 calcareous and non-calcareous sands and soils. The simplified method extracted equivalent quantities of the nonacid cations (data not shown,  $r^2 = 0.98$ , linear regression intercept and slope not significantly different from 0 and 1, respectively) when compared to the procedure as outlined by Houba et al. (2000).

All solutions were analyzed for nonacid cations using inductively-coupled plasma spectrometry. From the quantity of nonacid cations extracted by each method, we estimated the CEC by summation of the nonacid cations. We shall refer to this estimate of CEC as  $CEC_{sum}$ . The  $CEC_{sum}$  was calculated based on formulas modified from Sumner and Miller (1996) to report  $CEC_{sum}$  in units of mmol<sub>c</sub> kg<sup>-1</sup>:

 $CEC_{sum} = Ca + Mg + K + Na$ <sup>[1]</sup>

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where  $Ca = extracted Ca in mmol_c kg^{-1}$ 

Mg = extracted Mg in  $mmol_c kg^{-1}$ 

 $K = extracted K in mmol_c kg^{-1}$ 

 $Na = extracted Na in mmol_c kg^{-1}$ 

Thus, five  $\text{CEC}_{\text{sum}}$  estimates were obtained in addition to  $\text{CEC}_{\text{CE}}$  for each of the samples. Exchangeable acidity was measured for each sample with pH below 6 (Tables 6.1 and 6.2); we did not include the exchangeable acidity in our  $\text{CEC}_{\text{sum}}$  calculations.

Soil pH was measured in a 1:5 soil:water suspension. Calcium carbonate equivalent of each sample was measured using the method of Moore et al. (1987). Soil OM was measured by loss on ignition (Schulte, 1995). An empirical relationship based on the investigations of Helling et al. (1964) was used to calculate an expected contribution of organic matter to the sample CEC assuming a conversion factor from organic carbon to organic matter of 1.9:

$$CEC_{OM} (mmol_c kg^{-1}) = -311 + 268 \times pH$$
 [2]

The data were evaluated using least squares linear regression analysis. The  $CEC_{sum}$  values for each method were used to predict  $CEC_{CE}$ . All statistical analyses were performed using the REG procedure in SAS System Software version 9.1 (SAS, Cary, NC).

# **Results and Discussion**

## Sands

The extraction methods differed in their ability to predict  $CEC_{CE}$  of sand rootzones (Fig. 6.1). For the solutions that do not adjust to the sample pH during the extraction process (1 *M* NH<sub>4</sub>OAc, Mehlich 3, and Morgan), an overestimation of  $CEC_{CE}$  was observed in the sand samples with CCE > 10 g kg<sup>-1</sup>. This overestimation is due to the dissolution of solid-phase calcium or magnesium carbonates during the extraction process. Even when the samples with CCE > 10 g kg<sup>-1</sup> were omitted (Fig. 6.1), the 1 *M* NH<sub>4</sub>OAc, Mehlich 3, and Morgan tests produced  $CEC_{sum}$  values that overestimated  $CEC_{CE}$ .

The CEC<sub>OM</sub> estimates were of similar magnitude as CEC<sub>CE</sub> and were not influenced by CCE (Fig. 6.1). Because organic matter is the preponderant source of CEC in sand rootzones, one expects that OM and pH would predict CEC. The use of equation 2 provides a more accurate estimate of the CEC in sand rootzones than does CEC<sub>sum</sub> based on 1 *M* NH<sub>4</sub>OAc, Mehlich 3, or Morgan extractions.

Figure 6.1. The relationship between  $\text{CEC}_{\text{sum}}$  based on 1 *M* NH<sub>4</sub>OAc, Mehlich 3, Morgan, 1:5 H<sub>2</sub>O, and 0.01 *M* SrCl<sub>2</sub> extractions and corresponding  $\text{CEC}_{\text{CE}}$  for 37 sand samples varying in pH, CCE, and OM. The  $\text{CEC}_{\text{OM}}$  is also shown for comparison. The ( $\Box$ ) represent samples with CCE > 10 g kg<sup>-1</sup>, and the ( $\circ$ ) represent samples with CCE < 10 g kg<sup>-1</sup>.



The CEC<sub>sum</sub> derived from the 1:5  $H_2O$  extractions were always less than CEC<sub>CE</sub> (Fig. 6.1). The 1:5  $H_2O$  extraction removes only a fraction of the exchangeable nonacid cations, and although there is a positive relationship between CEC<sub>sum</sub> based on the 1:5  $H_2O$  extraction and CEC<sub>CE</sub>, it is possible that the sands with higher CEC also have a greater concentration of water soluble nonacid cations.

The 0.01 M SrCl<sub>2</sub> extraction resulted in CEC<sub>sum</sub> values that were comparable to CEC<sub>CE</sub> in all sands, regardless of CCE (Fig. 6.1). The 1 M NH<sub>4</sub>OAc, Mehlich 3, and Morgan procedures extract more nonacid cations, even in sands with low CCE, than would be exchangeable under simulated field conditions. However, the 0.01 M SrCl<sub>2</sub> method extracted nonacid cations of equivalent charge to those cations on the sample exchange sites.

All but one of the sands included in our analysis had  $CEC_{CE} < 60 \text{ mmol}_{c} \text{ kg}^{-1}$ . The recommended maximum organic matter content for sand rootzones maintained as creeping bentgrass [*Agrostis stolonifera* var. *palustris* (Huds.) Farw.] putting greens is 40 g kg<sup>-1</sup> (Carrow, 2003). At this maximum value of OM, and at a pH of 6.5, the  $CEC_{OM}$  (equation 2) would be 57 mmol<sub>c</sub> kg<sup>-1</sup>. Because turfgrass managers attempt to maintain the organic matter content of sand rootzones at less than 40 g kg<sup>-1</sup> (Carrow, 2003), most samples from sand rootzones probably fall within the linear prediction range of  $CEC_{CE}$  by the 0.01 *M* SrCl<sub>2</sub>  $CEC_{sum}$ .

Figure 6.2. The relationship between  $CEC_{sum}$  for 1 *M* NH<sub>4</sub>OAc, Mehlich 3, Morgan, 1:5 H<sub>2</sub>O, and 0.01 *M* SrCl<sub>2</sub> extractions and corresponding  $CEC_{CE}$  for 17 soil samples varying in pH, CCE, and OM. The  $CEC_{OM}$  is also shown for comparison. The ( $\Box$ ) represent samples with CCE > 10 g kg<sup>-1</sup>, and the ( $\circ$ ) represent samples with CCE < 10 g kg<sup>-1</sup>.



Soils

The CEC<sub>sum</sub> of soil samples extracted with 1 *M* NH<sub>4</sub>OAc, Mehlich 3, or Morgan solutions were of similar magnitude to CEC<sub>CE</sub>, particularly in soils with CCE  $< 10 \text{ g kg}^{-1}$  (Fig. 6.2). The CEC<sub>OM</sub>, which do not take into account the clay content of a soil, were less than CEC<sub>CE</sub> for these soil samples (Fig. 6.2). The 1:5 H<sub>2</sub>O CEC<sub>sum</sub> estimates were similar to the results from the sands in that they were unaffected by CCE but remained low in comparison with the CEC<sub>CE</sub> (Fig. 6.2). The predictive relationship between 0.01 *M* SrCl<sub>2</sub> CEC<sub>sum</sub> and CEC<sub>CE</sub> is linear with a slope of 1 up to about 60 mmol<sub>c</sub> kg<sup>-1</sup>, above which the 0.01 *M* SrCl<sub>2</sub> CEC<sub>sum</sub> underestimates CEC<sub>CE</sub> (Fig. 6.2).

#### **All Samples Combined**

Of the methods we evaluated for predicting  $CEC_{CE}$ , only 0.01 *M* SrCl<sub>2</sub> produces a suitable  $CEC_{sum}$  regardless of the sample CEC or CCE. The  $CEC_{sum}$  values based on this extraction had a quadratic predictive relationship (Fig. 6.3, A) with  $CEC_{CE}$  across all samples. For samples with  $CEC_{CE} > 60 \text{ mmol}_{c} \text{ kg}^{-1}$ , we measured the nonacid cations extracted by performing three consecutive 0.01 *M* SrCl<sub>2</sub> extractions on the same sample. When the  $CEC_{sum}$  from the first and second extractions were summed, the quadratic relationship between  $CEC_{sum}$  and  $CEC_{CE}$  became linear (Fig. 6.3, B). Three consecutive extractions with 0.01 *M* SrCl<sub>2</sub> produced a  $CEC_{sum}$  that overestimated  $CEC_{CE}$  (data not shown). The accuracy of the  $CEC_{sum}$  estimates could be slightly improved by subtracting the water soluble nonacid cations ( $CEC_{sum}$  derived from the 1:5 H<sub>2</sub>O extraction) from the 0.01 *M* SrCl<sub>2</sub> CEC<sub>sum</sub>, as shown in Fig. 6.3 (C).

Figure 6.3. The relationship between 0.01 *M* SrCl<sub>2</sub> CEC<sub>sum</sub> and CEC<sub>CE</sub> for 54 sand and soil samples varying in pH, CCE, and OM. The CEC<sub>sum</sub> is shown for one 0.01 *M* SrCl<sub>2</sub> extraction (A), for two consecutive 0.01 *M* SrCl<sub>2</sub> extractions (B), and for two consecutive 0.01 *M* SrCl<sub>2</sub> extractions with 1:5 H<sub>2</sub>O CEC<sub>sum</sub> subtracted (C). The ( $\Box$ ) represent samples with CCE > 10 g kg<sup>-1</sup>, and the ( $\circ$ ) represent samples with CCE < 10 g kg<sup>-1</sup>.



When the nonacid cations are represented as a percentage of the CEC, the implication is that the specified cation occupies the reported percentage of soil exchange sites. Because the aforementioned agronomic testing methods overestimate  $CEC_{CE}$ , it is not clear exactly what the nonacid cation percentage values then represent, other than simply a percentage of the cations extracted; it is misleading to consider them exchangeable for sand samples with  $CEC_{CE} < 60 \text{ mmol}_c \text{ kg}^{-1}$ .

We expect that a single 5 minute extraction with 0.01 *M* SrCl<sub>2</sub> will produce an accurate CEC estimate for most samples collected from sand rootzones or soils with  $CEC_{CE} < 60 \text{ mmol}_{e} \text{ kg}^{-1}$ , regardless of CCE. A notable exception is saline samples, from which no  $CEC_{sum}$ , regardless of extraction method, will provide an accurate estimate of CEC. It is not known if the 0.01 *M* SrCl<sub>2</sub> extraction results in accurate CEC estimates for sands to which certain types of inorganic or organic amendments have been added; our dataset did not include every possible soil amendment. It is furthermore unknown how accurate the method is in acid sands. However, turfgrass sand rootzones are usually limed to a pH of at least 5.5, so in practice it is unlikely to encounter samples with significant amounts of exchangeable Al. In addition to improved accuracy of the CEC estimates, a dilute salt solution that adjusts to the soil pH can be used to measure plant-available pools of soil P (Wendt and Corey, 1981) and soil micronutrients (McBride et al., 2003).

## Conclusion

Nonacid cations extracted by  $0.01 M \operatorname{SrCl}_2$  in a rapid 5 minute extraction procedure provided accurate estimates of  $\operatorname{CEC}_{CE}$  in sands and soils with  $\operatorname{CEC}_{CE}$  less than 60 mmol<sub>c</sub> kg<sup>-1</sup>, independent of the CCE of the samples. Because sand rootzones usually have a  $\operatorname{CEC}_{CE}$  less than 60 mmol<sub>c</sub> kg<sup>-1</sup>, use of a single 0.01 M SrCl<sub>2</sub> extraction will provide accurate estimates of exchangeable nonacid cations,  $CEC_{sum}$ , and resultant nonacid cation percentages. If  $CEC_{CE}$  is above 60 mmol<sub>c</sub> kg<sup>-1</sup>, two consecutive extractions are needed.

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