CHAPTER TWO

COMPARISON OF TWO METHODS FOR ESTIMATING PESTICIDE VOLATILIZATION FROM TURF

Abstract

Pesticide volatilization is a potential health hazard to turf users. Modeling pesticide volatilization can provide an effective tool for estimating environmental impact where complete information is unavailable. In the model developed (called Aero) the pesticide volatilization is directly related to the potential evapotranspiration, pesticide vapor pressure and the amount of pesticide left on the leaf surface. The latter was simulated as a first order pesticide degradation term and accounted for the rapid decrease in surface volatilization over time. The Aero model was tested against existing data from literature and compared to an existing model by Haith et al. (2002). The simulated volatilizations predictions were realistic and compared well with the observed data and the Haith model.

1. Introduction

Turfgrasses are plants that are used for a variety of purposes, including ornamentation, soil protection, and ground cover for recreational areas such as parks, golf courses, and athletic fields. Chemicals used for control of turfgrass pests may volatilize, resulting in potential health hazards for turf users (Clark *et al.*, 2000).

Pesticide volatilization is one of the dissipation processes affecting a wide variety of pesticides used in agriculture (Caro and Taylor, 1971; Cliath and Spencer, 1971; Farmer et al., 1972; Farmer et al. 1974; Harper et al., 1976; Spencer and

Farmer, 1980; Prueger *et al.*, 1999). Volatilization has been considered to be a major cause of pesticide loss from fields after application (Taylor *et al.*, 1977; Cooper *et al.*, 1990; Prueger *et al.*, 1999). Field studies have confirmed that pesticides lost from volatilization could range from negligible to 54% of the mass applied (Murphy *et al.*, 1996a, 1996b; Taylor *et al.*, 1977; Turner *et al.*, 1977). Most of the volatilization losses occur within the first few days of application and follow a diurnal pattern, with largest losses corresponding to high mid-day temperatures and solar radiation (Cooper, *et al.*, 1990; Haith *et al.*, 2002).

Due to the enormous number of pesticides currently in use, it is impractical to measure volatilization for each pesticide under different environmental conditions. Thus, effective mathematical models are needed that can predict the volatilization effects from pesticides where complete information on their impact on the environment is unavailable. Recently, Haith *et al.* (2002) presented such a model to predict pesticide volatilization from turf. Calibration using a specific pesticide was used as a method to develop the model to predict other independent data of different pesticides. The objective of this chapter was to develop a prediction model (Aero model) based upon a regression approach using the complete set of data used by Haith *et al.* (2002) to obtain pesticide degradation and absorption parameters in order to have a generalized prediction model of pesticide volatilization.

2. Model development

2.1 Aero model

A pesticide volatilization model is developed by drawing a parallel between water and pesticide vapor transport. The dispersal of residues into the atmosphere by vapor involves two distinct processes. The first is the volatilization of pesticide molecules into the air from the pesticide residue present on the soil or grass surfaces

and represents a phase change into vapor from the liquid or solid state. The vapor pressure of pesticide is a direct function of the concentration either in the water or as solid matter and can be described with Henry's law:

$$d = h \cdot C_p \tag{1}$$

where

d = vapor concentration of pesticide, mole/m³

 $C_p =$ solute concentration, mole/m³

h = Henry's law constant, dimensionless

Henry's law applies specifically to fluids but the same type of relationship holds for a solid when we assume that the rate of volatilization is equal to the pesticide density on the leaves.

The second process is the dispersion of the resulting pesticide vapor from turf surface into the overlying atmosphere by diffusion and turbulent transport (Taylor et al., 1990). This second process is the same one that controls the transfer of water vapor, carbon dioxide and other gasses between the turf surface and the overlying atmosphere. This dispersion process consists of molecular vapor diffusion through a thin laminar layer followed by turbulent transport. Turbulent transport may be visualized as macroscopic packets of air moving at random, there in convectively transporting their "cargoes" of energy, water and pesticide vapor from one spot to the next (Eagelson, 1970).

Process 1: Partition between air, water, and plant surfaces

Although the variety of chemicals among the pesticides will show different patterns in their physiochemical behavior in soils, many have similar patterns that are important in controlling their partition between the air, water, soil or plant surfaces. The partition of pesticide material between the plant surface and water phase depends on the partition between adsorption on the grass blade and the sorption and permeation into the cuticular waxes and the epidermal layer of the grass blade (Taylor and Spencer, 1990). The vapor density of the pesticide in the thin layer of air immediately in contact with the turf, to be referred to as the laminar boundary layer, is proportional to the pesticide solution concentration according to Equation 1. In order to relate this concentration to the amount applied, we introduce a factor "b" that represents the relationship between the vapor pressure in the air above the leaves and the amount of pesticide (either in dissolved or solid form) on the leaves. Thus, we can write:

$$C_p = \frac{P_A}{b} \tag{2}$$

where

 P_A = pesticide on plant leaves per unit area, mole m⁻²

Later on, we will define the b-term more closely. In this chapter, "b" will be used as a component of a fitting parameter; the units for "b" are m³ m⁻².

Process 2: Atmospheric transport

The flow of air over a turf surface is invariably turbulent except in a shallow layer with a depth of about 1 mm in which the flow may be regarded as laminar (Monteith, 1973). In this thin region of laminar flow, vapor transport is governed by molecular diffusion. Outside this zone in the overlying turbulent boundary layer, the dispersal of vapors, F, can be described in terms of an eddy diffusivity coefficient as (Taylor and Spencer 1990; Dingman 2002):

$$F = k_z \frac{dp}{dz}$$

where

 $F = vertical flux of vapor, mole m^{-2} sec^{-1}$

 k_z = mean eddy diffusivity coefficient or transport coefficient, m² sec⁻¹

 $p = vapor concentration, mole m^{-3}$

z = z coordinate in the vertical direction, m

Since Equation 3 is valid for turbulent transport, the molecular diffusion effects in the laminar flow layer are neglected (Taylor and Spencer, 1990). Since, both pesticide and water vapor transport from the turf layer through the laminar layer are similar molecular processes, the omission of molecular diffusion should not introduce a large error and in fact can be accounted for by using the eddy diffusivity approach. In turbulent transport theory, the eddy diffusion coefficient for water and pesticide vapor can be assumed to be the same, since it is the same transport phenomena governing both. Similar but in contrast, Jury and coworkers assumed that the molecular diffusion processes governed the pesticide release into a stagnant air layer immediately above the turf (Jury et al., 1983) and then proceeded to calculate the flux of pesticide into this stagnant boundary layer. The inherent assumption by Jury et al. being that what is transported by diffusion into the stagnant boundary layer will eventually be transported away by convection into the overlying atmosphere. Although the two approaches are conceptually the same in that they are both transport phenomena, the difference in approach is extremely important since the concentration gradient will be very different, as we will see later.

The equation for water vapor flux, F_w, becomes when averaged over sufficient long time that temporal variation can be neglected:

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(3)

$$F_{w} = -k_{x} \frac{e_{1} - e_{2}}{z_{1} - z_{2}} \tag{4}$$

where

 F_w = vapor flux of water, mole m⁻² hr⁻¹

 e_1 = vapor concentration of water at z_1 , mole m⁻³

 $e_2 =$ vapor concentration of water at z_2 , (mole m⁻³)

 z_1 = vertical direction at height 1, m

 z_2 = vertical direction at height 2, m

Similarly for pesticide vapor transport flux, F_p
$$F_p = -k_z \frac{d_1 - d_2}{z_1 - z_2}$$

where

 F_p = vapor flux of pesticide, mole m⁻² hr⁻¹

 d_1 = vapor concentration of pesticide at z_1 , mole/m³

 d_2 = vapor concentration of pesticide at z_2 , mole/m³

The subscripts 1 and 2 refer to the two different planes where the observations are made. In the following, we will take the plane with subscript 1 to be just above the turf grass surface and the second plane will be at the standard meteorological observation for vapor pressure measurement of 2 m. Because pesticides are applied only locally, we can take the pesticide concentration at this height equal to zero, i.e., $d_2 = 0$. By dividing Equation 4 and 5 and noting that the two eddy diffusion coefficients are assumed to be equal, we obtain:

$$F_{p} = F_{w} \frac{d_{1}}{e_{1} - e_{2}} \tag{6}$$

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(5)

By taking the first level (z_1) just above the turf surface, we can set the pesticide vapor concentration from Equation 1 equal to the d_1 in Equation 6. Moreover, since the turf is usually well watered we can also set e_1 equal to the saturated vapor concentration, e_s .

Then, if we were to assume that pesticide vapor is the only pesticide loss, we can write the amount of pesticide flux per unit area, $\frac{dP_A}{dt}$, as:

$$\frac{dP_A}{dt} = -F_w \frac{hC_p}{e_s - e_2} \tag{7}$$

where

 P_A = pesticide evaporation on a mass per unit area basis, mole m⁻²

Substituting the value of C_p in Equation 2 into Equation 7 gives:

$$\frac{dP_A}{dt} = -F_w \frac{h \cdot P_A}{b(e_s - e_2)} \tag{8}$$

Integrating Equation 8 and given at time 0, t = 0, $P_A = (P_A)_o$, then

$$P_{A} = (P_{A})_{o} \cdot \exp\left[-\left(\frac{F_{w}}{(e_{s} - e_{2})}\frac{h}{b}\right) \cdot t\right]$$
(9)

where

 $(P_A)_o$ = the initial mass of pesticide per unit area at time t = 0, mole m⁻²

The cumulative volatilized pesticide loss,
$$L_p$$
, can be expressed as:
 $L_p = (P_A)_o - P_A$ (10)

Substituting P_A in Equation 9 to Equation 10 and we obtain

$$L_{p} = (P_{A})_{o} \cdot \left[1 - \exp\left[\left(\frac{F_{w}}{(e_{s} - e_{2})} \frac{h}{b} \right) \cdot t \right] \right]$$
(11)

If the pesticide mass loss is disappearing by both volatilization (Equation 7) and also by degradation concurrently and degradation is occurring as a first order reaction, then Equation 9 could be modified

$$\frac{dP_A}{dt} = -\left(F_w \frac{h_b}{e_s - e_2} + a\right) P_A \tag{12}$$

where

 $a = degradation rate of pesticide, day^{-1}$

Integrating Equation 12 gives

$$P_{A} = (P_{A})_{o} \cdot \exp\left[-\left(\frac{F_{*}}{(e_{s} - e_{2})}\frac{h}{b} + a\right) \cdot t\right]$$
(13)

Development of Aero Model Constants

In the theoretical aerodynamic approach, there are two parameters which are not available from the literature. The first is the "b" term that includes the effect of adsorption and the second is the a-term term that includes the disappearance of the pesticide other than by volatilization. Also, Henry's constant is a variable during the whole experiment, since pesticides were mixed with water when sprayed onto the turfgrass (in liquid phase), but the pesticides may change to their solid phase (this means that a different value of Henry's constant would apply) due to lack of water on the surface of leaves. Instead of finding the b-term by itself, we determined the h/bterm as a single term instead of finding h and b terms separately to allow closure of the Aero model. We did this because both the h and b-terms are known for only certain temperature and phase value conditions; these values were not always known for the field data being used to develop the Aero model. These constants (a-term and h/b-term) will be determined for turfgrass by a regression analysis of observed pesticide volatilization data¹ (Haith *et al.*, 2002)

We have assumed that at each measurement, the amount of pesticide remaining on the leaves $((P_A)_i)$ could be expressed as:

$$(P_A)_{\tilde{t}} = (P_A)_o \cdot \exp(-a\tilde{t}) \tag{14}$$

where

 $(P_A)_{\tilde{t}}$ = amount of pesticide remaining on the leaves at time \tilde{t} , mole m⁻² \tilde{t} = time since initial pesticide spraying, day

Given the above assumption (Equation 14), pesticide volatilization loss could be described by combing Equation 8 and Equation 14:

$$\Delta(P_{A,v})_{i} = (P_{A})_{i} \cdot F_{w} \cdot \frac{h}{b} \cdot \frac{1}{e_{s} - e_{2}} \Delta t$$
(15)

where

 $\Delta(P_{A,v})_{\tilde{t}}$ = pesticide loss (volatilization) at time \tilde{t} in short period of time Δt Δt = sampling period of pesticide volatilization, day

Further rearranging Equation 15, we can obtain

$$\Delta(P_{A,v})_{\bar{t}} = (P_A)_o \cdot \exp\left(-a\bar{t}\right) \cdot F_w \cdot \frac{h}{b} \cdot \frac{1}{e_s - e_2} \Delta t \tag{16}$$

¹ Haith *et al.* (2002) provides most of the data used in this analysis to develop the Aero model. Some data may not be explicitly shown in Haith *et al.* (2002) and for these cases, this data is shown either in the Tables of this Chapter or the Chapter Appendix; the original data were collected at the University of Massachusetts Turfgrass Research Center in South Deerfield, MA. When the text of this Chapter refers to Haith *et al.* (2002), this infers that it may be referring to the University of Massachusetts data.

By rearranging Equation 16, we obtain

$$\frac{\frac{\Delta(P_{A,v})_{\tilde{t}}}{(P_{A})_{o}}}{F_{w} \cdot \frac{1}{e_{s} - e_{2}} \Delta t} = \exp\left(-a\tilde{t}\right) \cdot \frac{h}{b}.$$
(17)

By taking the natural log of both sides of the equal sign in Equation 17, we obtain

$$\ln \frac{\frac{\Delta(P_{A,v})_{\tilde{t}}}{(P_{A})_{o}}}{F_{w} \cdot \frac{1}{e_{s} - e_{2}} \Delta t} = -a\tilde{t} + \ln \frac{h}{b}.$$
(18)

Since $\Delta(P_{A,v})_{\tilde{t}}$, $(P_A)_o$, F_w , e_s , e_2 and Δt are accessible values, we can obtain a and $\frac{h}{b}$ by

plotting $\ln \frac{\frac{\Delta(P_{A,\nu})_{\tilde{t}}}{(P_A)_o}}{F_{\nu} \cdot \frac{1}{e_s - e_2} \Delta t}$ versus \tilde{t} from the regression line. Note that $\Delta(P_{A,\nu})_{\tilde{t}}$ here is

the observed pesticide volatilization. The values for $\Delta(P_{A,\nu})_{\bar{i}}$, $(P_A)_o$, Δt and the procedure to calculate F_w , e_s , e_2 are described by Haith *et al.* (2002).

Once a term and $\frac{h}{b}$ term were determined, we can insert these two values into Equation 16. This then becomes the Aero model and can be used as a general prediction equation to predict pesticide volatilization.

In order to compare observed and predicted data from field studies as given by Haith *et al.* (2002), it was necessary to change the units used in the Aero model developed above. Since the unit of $\Delta(P_{A,\nu})_i$ in Equation 16 is mole m⁻², the units were changed to g ha⁻¹ as used in Haith *et al.* (2002) paper. By multiplying $\Delta(P_{A,\nu})_i$ with (10,000 M_w), we obtain $\Delta(P_{A,\nu})_i$ in the same units of g ha⁻¹, where M_w is the molecular weight of the particular pesticide.

2.2 Haith Model

The Haith *et al.* (2002) model (to be referred to as the Haith model from here on out) was developed by linking pesticide volatilization rates to evapotranspiration values, as determined by the Penman equation (Jensen et al., 1990):

$$\Delta(\mathbf{P}_{A,H})_{\tilde{t}} = k \cdot \left(F_{W,H} \cdot \Delta t\right) \cdot \left(\frac{d_s}{e_s}\right) \cdot \left(\frac{\lambda_{w,t}}{\lambda_{p,t}}\right) \cdot \left(P_{A,H}\right)_t$$
(19)

$$(P_{A,H})_{t+1} = \cdot (P_{A,H})_t \cdot e^{-\alpha \cdot t'} - \Delta (\mathbf{P}_{A,H})_{\tilde{t}}$$

$$(20)$$

where

 $\Delta(P_{A,H})_i$ = pesticide vaporized from surface vegetation during time Δt at time step t of Haith model, g ha⁻¹

k = volatilization constant, mm⁻¹

 $F_{w,H} \Delta t$ = water evapotranspiration during time Δt , mm

 d_s = saturated pesticide vapor pressure, the same unit as e_s , kPa or mole m⁻³

 $\lambda_{p,t}$ = latent heat of vaporization of the chemical during time Δt at time step t, J g⁻¹ or kJ kg⁻¹

 $\lambda_{w,t}$ = latent heat of vaporization of water during time Δt at time step t, J g⁻¹ or kJ kg⁻¹ (P_{A,H})_t = pesticide available for volatilization on vegetation at the beginning time step t, g ha⁻¹ $(P_{A,H})_{t+1}$ = pesticide available for volatilization on vegetation at the beginning of time step t+1, g ha⁻¹

 α = degradation rate of pesticide on vegetation surface, hr⁻¹

t' = time elapsed between successive sampling periods, hr

Note that t' is the time elapsed between successive sampling periods. Note that Δt is usually only a portion of t', e.g. t' might be 6 hours or 24 hours and Δt is usually around 2 to 4 hours of sampling time. Only the Haith model uses the variable t'.

The evapotranspiration value as shown in Equation 19 was adjusted based upon the ratios of water and chemical saturated vapor pressures (d_s / e_s) and latent heats of vaporization ($\lambda_{w,t}/\lambda_{p,t}$) to predict pesticide vaporization (Haith et al., 2002). The pesticide chemicals or simply pesticides are assumed to have first order degradation on turf grass over time. Field data as used by Haith *et al.* (2002) for eight pesticides was available to use in model comparisons made between the Aero and the Haith models. The eight pesticides were Bendiocarb, Diazinon, Ethoprop, Isazofos, Carbaryl, Chlorpyrifos, Isofenphos and Trichlorfon. These eight pesticides were classified into being either high volatilization or low volatilization pesticides, based upon their pesticide saturation vapor pressure (Haith *et al.*, 2002).

Haith *et al.* (2002) found by calibration that $k = 130 \text{ mm}^{-1}$ for high volatilization pesticide (classified as Group 1 in Haith *et al.*, 2002) and 405 mm⁻¹ for low volatilization pesticide (classified as Group 2 in Haith *et al.*, 2002)

All the chemical characteristics and calculations related to $F_{w,H}$, d_s , e_s , $\lambda_{w,t}$, $\lambda_{p,t}$ and C_t are described by Haith *et al.*, 2002. Values used in the simulation analysis for saturated vapor pressure, molecular weight, solubility, Henry's constant and K_{oc} are shown in Table 2.1.

	K_{oc} , cm ³ g ⁻¹ ,	Saturated	**	** Solid or	**	** Henry's
	from Haith	Vapor	Molecular	Liquid at	Solubility	Constant (h),
	et al., 2002	Pressure at	Weight, g	room	in Water,	dimensionless
		25 °C, from	mole ⁻¹	temperature	ppm	
		Haith et al.,				
		2002, kPa				
Bendiocarb	385	4.6E-06	223.2	Solid	40~260	9.03E-07
					@20°C	
Diazinon	1520	1.2E-05	304.3	Liquid	40~68 @20	1.53E-05
					°C	
Ethoprop	104	4.3E-05	242.3	Liquid	700~750	3.78E-06
					@20 °C	
Isazofos	155	1.2E-05	313.7	Liquid	34~69 @	5.71E-06
					20 °C	
Carbaryl	288	4.9E-08	201.2	Solid	~105 @ 20	1.86E-08
					°C	
Chlorpyrifos	9930	2.7E-06	350.6	Solid	1.18 @ 25	1.93E-04
					°C	
Isofenphos	777	4.4E-07	345.4	Liquid	18~23 @	1.73E-06
					20 °C	
Trichlorfon	15	5.0E-07	257.4	Solid	130000 @	2.54E-10
					20 °C	

Table. 2.1. Characteristics of pesticides used in this modeling effort.

** Agricultural Research Service, USDA database, 2001.

2.3 Parameter Adjusting for Haith Model and Aero Model

The Haith model, $\Delta(P_{A,H})_i$, (Equation 19) and Aero model, $\Delta(P_{A,v})_i$, (Equation 16) were compared by predicting the same observed data set as used by Haith *et al.* (2002). The input parameters used by the Haith model $\Delta(P_{A,H})_i$ include $F_{w,H} \Delta t$, k, d_s, e_s, $\lambda_{p,t}$, $\lambda_{w,t}$, α and t'. The input parameters used in the Aero model to predict $\Delta(P_{A,v})_i$ include (P_A)₀, $F_w\Delta t$, h/b, e_s, e₂ and Δt . The methods used to develop the closure constants for the Aero model are discussed below.

2.3.1 $F_{w,H}$ Δt and $F_w \Delta t$:

Water evaporation during time Δt in the Haith model, $E_w \Delta t$, is in units of mm and E_w is calculated from the Penman equation as described in Haith *et al.*, 2002 and described by Jensen *et al.*, (1990). The detailed equations are given in Appendix 2.3. The input parameters to obtain $F_{w,H} \Delta t$ such as air temperature, solar radiation, wind speed, duration time (Δt) and lowest temperature of a day are listed on Appendix 2.1.

Water evaporation during time Δt in the Aero model, $F_w \cdot \Delta t$, is in units of mole m⁻². These units can be converted to the same units used to describe $F_{w,H} \cdot \Delta t$ of mm as follows:

$$F_{w} \cdot \Delta t = F_{w,H} \cdot \Delta t \cdot \frac{1000}{18}$$
⁽²¹⁾

2.3.2 e_s and e_2

Saturated water vapor pressure can be directly related to air temperature as described by Jensen et al., (1990):

 $e'_{s} = 3.38639[(0.00738 T + 0.8072)^{8} - 0.000019|1.8 T + 48|+0.001316]$ (22) where

 e'_{s} = saturated water vapor pressure at air temperature T, kPa

T = air temperature during time Δt , °C

The units of e'_s are kPa and need to be converted to units of mole m⁻³ as is used in the Aero model, e_s (Equation 16). The relationship between e'_s and e_s can be obtained by using the ideal gas law (see Appendix 2.4):

$$\mathbf{e}_{s} = \mathbf{e}'_{s} \cdot \frac{1000}{R \cdot T_{a}} \tag{23}$$

where

R = gas constant, 8.314 J mole⁻¹ K⁻¹

 T_a = air temperature during time Δt , K

The actural water vapor pressure, e'_2 (units of kPa), can be calculated as:

 $e'_2 = e'_s \cdot RH \tag{24}$

where

RH = relative humidity during time Δt , dimensionless

Since there was no RH data recorded during the experiments, the alternative method, addressed in Haith (2002), to estimate water vapor pressure is to use the dew point temperature (T_d) for the temperature value of T in Equation 22. The dew point temperature is estimated as the lowest temperature of the day (Timmons and Gates, 1986). Once e'₂ is calculated, then e₂ (in unit of mole m⁻³) can be calculated by using Equation 23 to convert to units of mole m⁻³.

2.3.3 ds,

Calculation of the saturated vapor pressure of pesticide at temperature T, d_s, in the Haith model (Equation 19) can be described as (Haith *et al.*, 2002, Grain, 1982):

$$\ln\left(\frac{\mathrm{d}_{\mathrm{s}}}{\mathrm{d}_{\mathrm{s},T_{\mathrm{a}_{0}}}}\right) = \left[\frac{\mathrm{M}_{\mathrm{w}}\cdot\lambda_{\mathrm{p},\mathrm{T}_{\mathrm{a}_{0}}}}{\Delta Zb\cdot\mathrm{R}\cdot\mathrm{T}_{\mathrm{a}_{0}}}\right] \cdot \left(1 - \left(3 - 2\cdot\frac{\mathrm{T}_{\mathrm{a}}}{T_{a_{0}}}\right)^{m} \cdot\frac{T_{a_{0}}}{T_{a}} \cdot -2m\left(3 - 2\cdot\frac{\mathrm{T}_{\mathrm{a}}}{T_{a_{0}}}\right)^{m-1} \cdot\ln\frac{\mathrm{T}_{\mathrm{a}}}{T_{a_{0}}}\right)$$
(25)

where $d_{s,T_{a_0}} =$ saturated pesticide vapor pressure at temperature T_{a_0} , kPa $T_{a_0} =$ temperature at the addressed $d_{s,T_{a_0}}$, K $\lambda_{p,T_{a_0}} =$ latent heat of pesticide vaporization at temperature T_{a_0} , J/g $\Delta Z_b =$ compressibility factor at boiling point, dimensionless, 0.97 (Grain, 1982) m = constant, 0.8 (Haith *et al.*, 2002)

The data for saturated vapor pressure of each tested pesticide at 25 °C are shown on Table 2.1.

2.3.4 $\lambda_{w,t}, \lambda_{p,t}$

The latent heat of vaporization of water $(\lambda_{w,t})$ and pesticide $(\lambda_{p,t})$ used in the Haith model (Equation 19) can be described as (Haith *et al.*, 2002, Jensen 1990)

 $\lambda_{w,t} = 2501 - 2.361 T_t$

where

 T_t = temperature during time Δt , °C

The pesticide latent heat of vaporization $(\lambda_{p,t})$ used in the Haith model (Equation 19) can be estimated as (Haith, 2002; Grain, 1982):

$$\frac{M_{w} \cdot \lambda_{P,T_{a_{0}}}}{R \cdot T_{a_{0}}} = K_{f} \cdot \left(9.03 + \ln \frac{T_{a_{0}}}{d_{s,T_{a_{0}}}}\right)$$
(27)

where

Kf = constant, dimensionless, 1.06 (Haith et al., 2002)

For different temperatures, e.g. T_a , the pesticide latent heat of vaporization could also be estimated by using the same equation (Equation 27) and substituting T_{a_0} with T_a .

2.3.5 Initial Pesticide Applied

The initial amount of pesticide applied for each experiment is shown in Appendix 2.2. Note that the units for the initial amount of pesticide in Haith *et al.*, 2002 $((P_{A,H})_0)$ is g ha⁻¹ and the units used in the Aero model $((P_A)_o)$ is mole m⁻². The conversion of these units could be described as:

$$(P_A)_0 = \frac{(P_{A,H})_0}{10000 \cdot M_w}$$
(28)

2.4 Model Comparison

Both the Aero and Haith models were used to predict pesticide volatilization for each period (Δt) within a data set for a particular pesticide. The cumulative pesticide volatilization for a particular pesticide (sum of the mass volatization for all the periods within a set of experiments for a particular pesticide) were also calculated and compared for both two models. Comparisons between the two models was facilitated by forcing the Aero model to predict volatilization in the same units as used by the Haith model.

2.5 Calculation of Half Life

The half life can be calculated as:

$$\Delta T_{hf} = \frac{0.693}{\alpha}$$

where

 ΔT_{hf} = half life of pesticide calculated from a, day

(29)

Results and Discussion

The graphs of $\frac{\frac{\Delta(P_A)_{\bar{t}}}{(P_A)_o}}{F_w \cdot \frac{1}{e_x - e_2} \Delta t}$ versus \bar{t} of each pesticide are shown in Figures

2.1–2.8. From these graphs, and by taking the natural log of $\frac{\Delta(P_A)_{\tilde{i}}}{(P_A)_o}$, we obtain $F_* \cdot \frac{1}{e_s - e_2} \Delta t$

the a-term and h/b-term directly from the equation regression line (fitted curve) when done in log form. Since h (Henry's constant) is different when pesticide is either in liquid phase (at the first beginning of pesticide applied) or solid phase (after pesticide applied for a while), we represent h/b instead of b to address this issue. The a-term and h/b term of each pesticide as developed for the Aero model as just described are given in Table 2.2. It is customary to express the a-term as a half life. The relationship between a and half life is shown in Equation 29. The half life calculated from the a-term term ranged from 0.8 to 3.2 days.

Comparing to the half life values from USDA database (shown on Table 2.13), the half life calculated from the a-term term is relatively small. Since there are several forms of degradation, such as hydrolysis, photolysis, bio-degradation, then each of these forms of degradation could be represented by a different half life for the same pesticide. The half life of hydrolysis, photolysis and field dissipation also shown on Table 2.13. Also, the a-term term in the Aero model effectively lumped all degradation forms into one term; it also indirectly included any pesticide dissipation due to wash off. Thus, it is reasonable that the pesticide half life represented by the aterm is smaller than the half life from other individual sources.

Using the a-term and h/b-term which are uniquely determined for each of the eight pesticides modeled as given in Table 2.2 and inserting them into the Aero model,

predictions are made and are given in Tables 2.3-2.10 along with the observed values and the Haith model predictions. The data is presented in all cases in Tables 2.3–2.10 as a percentage of total pesticide applied. A graphical comparison is shown in Figures 2.9–2.16 of the two prediction models versus observed data where a perfect fit would be 45°-line. Similarly, the observed cumulative sum of pesticide volatilization for each experiment versus both model's predicted volatilization are shown in Table 2.11.

The R^2 values for each model to predict the observed pesticide volatilization data are shown in Table 2.12. There are two different R^2 in this table. One is to compare each period of modeled volatilization with observed values (R^2 , individually) and the other is to compare the sum of the quantity of pesticide volatized for each period over the entire experiment (R^2 , sum). For the individual period predictions, the Aero model compared to the Haith model R^2 is larger in seven out of the eight pesticides. In comparing the cumulative sum of pesticide volatilization, both models appear to be similar as both models had the same quantity of larger R^2 values.

If R^2 is used as the tool to judge which model is superior, then the R^2 calculated from Aero model and observed values of most of the pesticide is greater than that calculated from Haith model. Thus, one could conclude that the Aero model is a better predictor of pesticide volatilization than the Haith model. One might expect that the Aero model should exhibit higher R^2 values than the Haith model for thie individual periods, since the Aero model is based upon the data which is then predicted. However, the Aero model in contrast to the Haith model does not require classifying the pesticides into a class of either high or low volatilization potential. Thus, one could argue that the Aero model is a more general model for predicting pesticide volatilization behavior. The Aero model has also introduced a different form of the degradation term. Additional pesticide data is needed to fully evaluate the comparisons between these two models.

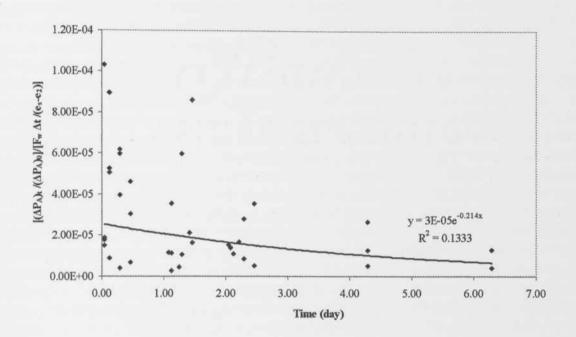


Figure 2.1. The graph of $\frac{\frac{\Delta(P_A)_{\tilde{t}}}{(P_A)_o}}{F_w \cdot \frac{1}{e_s - e_2} \Delta t}$ versus time of Bendiocarb.

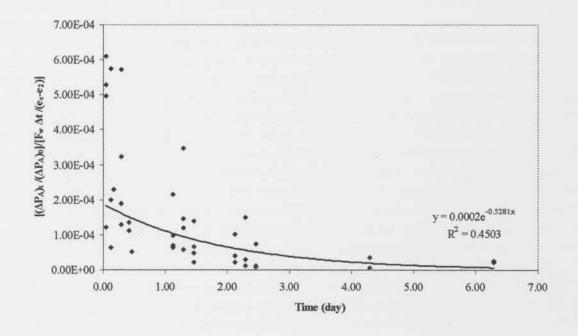
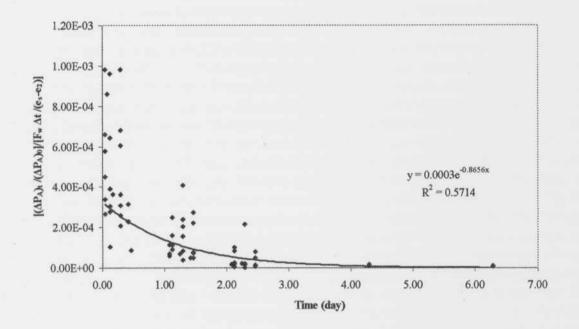
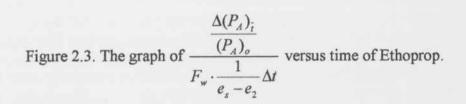
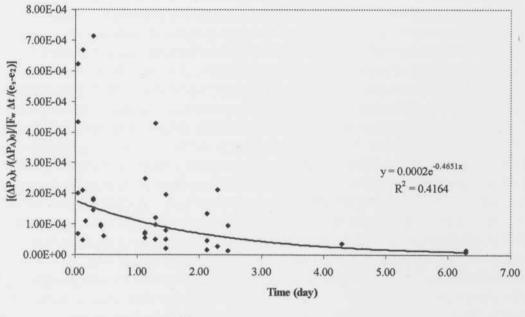
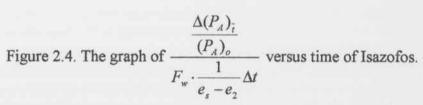


Figure 2.2. The graph of $\frac{\frac{\Delta(P_A)_{\tilde{t}}}{(P_A)_o}}{F_w \cdot \frac{1}{e_s - e_2} \Delta t}$ versus time of Diazinon.









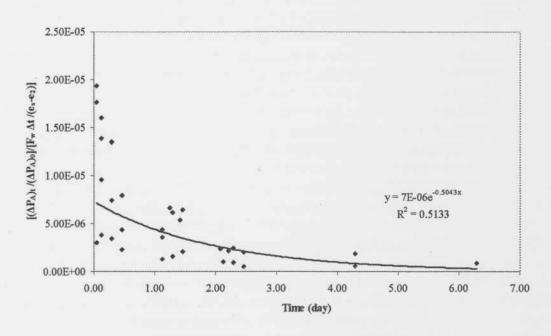
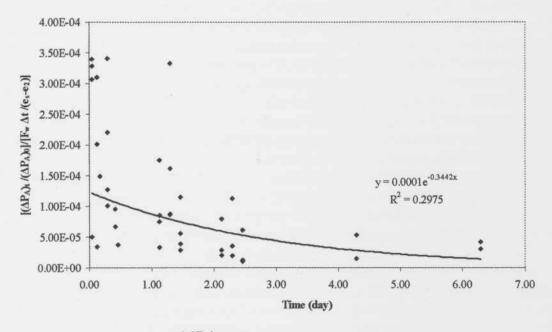
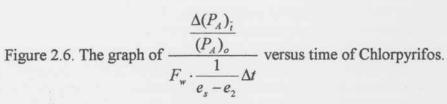
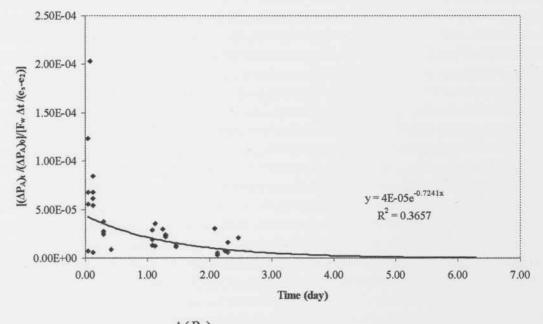
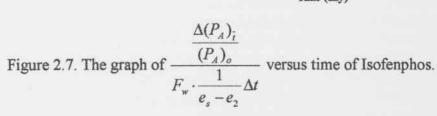


Figure 2.5. The graph of $\frac{\frac{\Delta(P_A)_{\tilde{t}}}{(P_A)_o}}{F_{w} \cdot \frac{1}{e_s - e_2} \Delta t}$ versus time of Carbaryl.









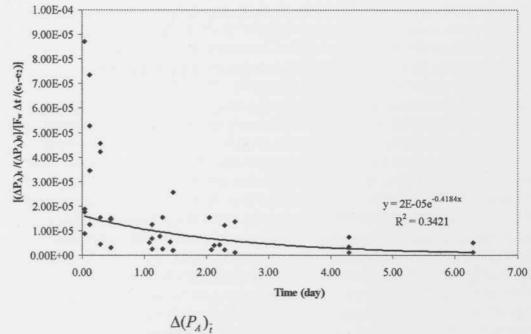


Figure 2.8. The graph of $\frac{\frac{\Delta(P_A)_{\tilde{t}}}{(P_A)_o}}{F_w \cdot \frac{1}{e_s - e_2} \Delta t}$ versus time of Trichlorfon.

allary SIS.			
	a (day ⁻¹)	Half Life fro	m a h/b (m^3/m^2)
		(day)	
Bendiocarb	0.21	3.24	2.56E-05
Diazinon	0.53	1.31	1.88E-04
Ethoprop	0.87	0.80	3.28E-04
Isazofos	0.47	1.49	1.77E-04
Carbaryl	0.50	1.37	7.25E-06
Chlorpyrifos	0.34	2.01	1.23E-04
Isofenphos	0.72	0.96	4.37E-05
Trichlorfon	0.42	1.66	1.61E-05

Table 2.2. The a-term and h/b-term of each pesticide as determined by regression analysis.

Table 2.3. The comparison of Haith et.al model and Aero model of Bendiocarb. (High volatilization group)

Exp	Exp	Exp. #	Time	Observed	Haith	Aero
teach.	P		(day)	(%)	(%)	(%)
Week 3	95-2	D1-V1	0.04	0.32	0.14	
WOOR 5		D1-V2	0.13	0.45		
		D1-V3	0.29			
		D2-V1	1.13	0.13		
		D2-V2	1.25			
		D2-V3	1.42	0.04		0.04
		D3-V1	2.08			
		D3-V2	2.21	0.08		
		D5-V1	4.29			0.10
		D7-V1	6.29			
Week 6	95-4	D1-V1	0.04			
		D1-V2	0.13			
		D1-V3	0.29			
		D1-V4	0.46			0.18
		D2-V1	1.13			
		D2-V2	1.29			
		D2-V3	1.46			
		D3-V1	2.04	0.04	0.04	0.04
		D3-V2	2.29	0.23	0.24	0.13
		D3-V3	2.46	0.18	0.08	0.08
		D5-V1	4.29	0.29	0.25	0.11
		D7-V1	6.29	0.15	0.14	0.08
Week 9	95-6	D1-V1	0.04	0.02	0.01	0.03
		D1-V2	0.13	0.11	0.01	0.05
		D1-V3	0.29	0.23	0.04	0.09
		D1-V4	0.46	0.14	0.05	0.11
		D2-V1	1.08	0.09	0.17	0.16
Week 13	96-2	D1-V1(rain	i) 0.04	0.05	0.03	0.09
		D1-V2	0.13	0.07	0.17	0.20
		D1-V3(rain	i) 0.29	0.07	0.53	0.42
		D1-V4(rair	i) 0.46	0.09	0.27	0.30
		D2-V1(rain	1.13	0.04	0.16	0.31
		D2-V2(rain	i) 1.29	0.20	0.42	0.37
		D2-V3(rain	i) 1.46	0.16	0.14	0.18
		D3-V1	2.13	0.22	0.05	0.33
		D3-V2	2.29	0.19	0.30	0.35
		D3-V3(rain	i) 2.46	0.10	0.10	0.29
		D5-V1(rain	n) 4.29	0.22	0.17	0.18
R ²					0.351	0.013

 \mathbb{R}^2

0.351 0.013

Table 2.4. The comparison of Haith et.al model and Aero model of Diazinon.

(High volatilization group)

D1-V1 D1-V2 D1-V3 D2-V1 D2-V2 D2-V3 D3-V1 D3-V2 D3-V3 D5-V1 D7-V1 D1-V1 D1-V2 D1-V3 D1-V4 D2-V1 D2-V2 D2-V3 D3-V2 D3-V3 D3-V2 D3-V3 D5-V1	(day) 0.04 0.13 0.29 1.13 1.29 1.46 2.13 2.29 2.46 4.29 0.04 0.17 0.29 0.04 0.17 0.29 0.42 1.13 1.29 1.46 2.29 2.46 4.29 0.42 1.13 1.29 0.44 0.17 0.29 0.44 0.17 0.29 0.44 0.17 0.29 0.44 0.17 0.29 0.44 0.13 0.29 0.46 1.13 1.29 1.46 2.13 0.29 0.46 1.13 1.29 0.46 1.13 1.29 1.46 1.29 0.46 1.13 1.29 0.46 1.13 1.29 0.46 1.13 1.29 0.46 0.13 0.29 0.46 0.13 0.29 0.42 1.13 1.29 0.42 1.13 1.29 0.42 1.13 1.29 0.42 1.13 1.29 0.42 1.13 1.29 0.42 1.46 0.29 0.42 1.13 1.29 0.42 1.13 1.29 0.42 1.13 1.29 1.46 0.29 0.42 1.13 1.29 1.46 0.42 0.42 1.13 1.29 1.46 2.29 0.42 1.13 1.29 1.46 2.29 0.42 1.43 1.29 1.46 2.29 1.46 2.29 0.42 1.43 1.29 1.46 2.29 1.46 2.29 1.46 2.29 1.46 2.29 1.46 2.29 1.46 2.29 1.46 2.29 1.46 2.29 1.46 2.29 1.46 2.29 1.46 2.29 2.46 2.29 1.46 2.29 2.46 4.29 1.46 2.29 2.46 4.29 1.46 2.29 2.46 4.29	(%) 1.08 1.35 2.22 3.88 4.30 0.45 0.86 1.53 0.72 0.37 0.26 0.42 1.47 3.29 0.68 0.66 1.37 0.35 0.33	(%) 0.08 0.13 0.17 0.74 1.54 0.12 1.14 2.62 0.68 2.29 2.18 0.02 0.70 2.51 0.43 1.03 2.75 0.93 1.98	(%) 0.33 0.41 0.62 1.86 1.18 0.28 0.52 0.57 0.51 0.20 0.07 0.16 1.10 1.64 0.76 0.70 0.89 0.65
D1-V2 D1-V3 D2-V1 D2-V2 D2-V3 D3-V1 D3-V2 D3-V3 D5-V1 D7-V1 D1-V2 D1-V3 D1-V4 D2-V1 D2-V2 D2-V3 D3-V2 D3-V2 D3-V3	0.13 0.29 1.13 1.29 1.46 2.13 2.29 2.46 4.29 6.29 0.04 0.17 0.29 0.42 1.13 1.29 1.46 2.29 2.46	$\begin{array}{c} 1.35\\ 2.22\\ 3.88\\ 4.30\\ 0.45\\ 0.86\\ 1.53\\ 0.72\\ 0.37\\ 0.26\\ 0.42\\ 1.47\\ 3.29\\ 0.68\\ 0.66\\ 1.37\\ 0.35\\ 0.33\end{array}$	0.13 0.17 0.74 1.54 0.12 1.14 2.62 0.68 2.29 2.18 0.02 0.70 2.51 0.43 1.03 2.75 0.93	0.41 0.62 1.86 1.18 0.28 0.52 0.57 0.51 0.20 0.07 0.16 1.10 1.64 0.76 0.70 0.89
D1-V3 D2-V1 D2-V2 D2-V3 D3-V1 D3-V2 D3-V3 D5-V1 D7-V1 D1-V2 D1-V3 D1-V4 D2-V1 D2-V2 D2-V3 D3-V2 D3-V2 D3-V3	0.29 1.13 1.29 1.46 2.13 2.29 2.46 4.29 6.29 0.04 0.17 0.29 0.42 1.13 1.29 1.46 2.29 2.46	$\begin{array}{c} 2.22\\ 3.88\\ 4.30\\ 0.45\\ 0.86\\ 1.53\\ 0.72\\ 0.37\\ 0.26\\ 0.42\\ 1.47\\ 3.29\\ 0.68\\ 0.66\\ 1.37\\ 0.35\\ 0.33\end{array}$	0.17 0.74 1.54 0.12 1.14 2.62 0.68 2.29 2.18 0.02 0.70 2.51 0.43 1.03 2.75 0.93	0.62 1.86 1.18 0.28 0.52 0.57 0.51 0.20 0.07 0.16 1.10 1.64 0.76 0.70 0.89
D2-V1 D2-V2 D2-V3 D3-V1 D3-V2 D3-V3 D5-V1 D7-V1 D1-V2 D1-V3 D1-V4 D2-V1 D2-V2 D2-V3 D3-V2 D3-V3	1.13 1.29 1.46 2.13 2.29 2.46 4.29 6.29 0.04 0.17 0.29 0.42 1.13 1.29 1.46 2.29 2.46	3.88 4.30 0.45 0.86 1.53 0.72 0.37 0.26 0.42 1.47 3.29 0.68 0.66 1.37 0.35 0.33	0.74 1.54 0.12 1.14 2.62 0.68 2.29 2.18 0.02 0.70 2.51 0.43 1.03 2.75 0.93	$\begin{array}{c} 1.86\\ 1.18\\ 0.28\\ 0.52\\ 0.57\\ 0.51\\ 0.20\\ 0.07\\ 0.16\\ 1.10\\ 1.64\\ 0.76\\ 0.70\\ 0.89\end{array}$
D2-V2 D2-V3 D3-V1 D3-V2 D3-V3 D5-V1 D7-V1 D1-V1 D1-V2 D1-V3 D1-V4 D2-V1 D2-V2 D2-V3 D3-V2 D3-V3	1.29 1.46 2.13 2.29 2.46 4.29 6.29 0.04 0.17 0.29 0.42 1.13 1.29 1.46 2.29 2.46	$\begin{array}{c} 4.30\\ 0.45\\ 0.86\\ 1.53\\ 0.72\\ 0.37\\ 0.26\\ 0.42\\ 1.47\\ 3.29\\ 0.68\\ 0.66\\ 1.37\\ 0.35\\ 0.33\end{array}$	1.54 0.12 1.14 2.62 0.68 2.29 2.18 0.02 0.70 2.51 0.43 1.03 2.75 0.93	$\begin{array}{c} 1.18\\ 0.28\\ 0.52\\ 0.57\\ 0.51\\ 0.20\\ 0.07\\ 0.16\\ 1.10\\ 1.64\\ 0.76\\ 0.70\\ 0.89\end{array}$
D2-V3 D3-V1 D3-V2 D3-V3 D5-V1 D7-V1 D1-V1 D1-V2 D1-V3 D1-V4 D2-V1 D2-V2 D2-V3 D3-V2 D3-V3	1.46 2.13 2.29 2.46 4.29 6.29 0.04 0.17 0.29 0.42 1.13 1.29 1.46 2.29 2.46	0.45 0.86 1.53 0.72 0.37 0.26 0.42 1.47 3.29 0.68 0.66 1.37 0.35 0.33	0.12 1.14 2.62 0.68 2.29 2.18 0.02 0.70 2.51 0.43 1.03 2.75 0.93	0.28 0.52 0.57 0.51 0.20 0.07 0.16 1.10 1.64 0.76 0.70 0.89
D3-V1 D3-V2 D3-V3 D5-V1 D7-V1 D1-V1 D1-V2 D1-V3 D1-V4 D2-V1 D2-V2 D2-V3 D3-V2 D3-V3	2.13 2.29 2.46 4.29 6.29 0.04 0.17 0.29 0.42 1.13 1.29 1.46 2.29 2.46	0.86 1.53 0.72 0.37 0.26 0.42 1.47 3.29 0.68 0.66 1.37 0.35 0.33	1.14 2.62 0.68 2.29 2.18 0.02 0.70 2.51 0.43 1.03 2.75 0.93	0.52 0.57 0.51 0.20 0.07 0.16 1.10 1.64 0.76 0.70 0.89
D3-V2 D3-V3 D5-V1 D7-V1 D1-V2 D1-V2 D1-V3 D1-V4 D2-V1 D2-V2 D2-V3 D3-V2 D3-V3	2.29 2.46 4.29 6.29 0.04 0.17 0.29 0.42 1.13 1.29 1.46 2.29 2.46	$\begin{array}{c} 1.53\\ 0.72\\ 0.37\\ 0.26\\ 0.42\\ 1.47\\ 3.29\\ 0.68\\ 0.66\\ 1.37\\ 0.35\\ 0.33\end{array}$	2.62 0.68 2.29 2.18 0.02 0.70 2.51 0.43 1.03 2.75 0.93	0.57 0.51 0.20 0.07 0.16 1.10 1.64 0.76 0.70 0.89
D3-V3 D5-V1 D7-V1 D1-V2 D1-V2 D1-V3 D1-V4 D2-V1 D2-V2 D2-V3 D3-V2 D3-V3	2.46 4.29 6.29 0.04 0.17 0.29 0.42 1.13 1.29 1.46 2.29 2.46	0.72 0.37 0.26 0.42 1.47 3.29 0.68 0.66 1.37 0.35 0.33	0.68 2.29 2.18 0.02 0.70 2.51 0.43 1.03 2.75 0.93	0.51 0.20 0.07 0.16 1.10 1.64 0.76 0.70 0.89
D5-V1 D7-V1 D1-V2 D1-V2 D1-V3 D1-V4 D2-V1 D2-V2 D2-V3 D3-V2 D3-V3	4.29 6.29 0.04 0.17 0.29 0.42 1.13 1.29 1.46 2.29 2.46	0.37 0.26 0.42 1.47 3.29 0.68 0.66 1.37 0.35 0.33	2.29 2.18 0.02 0.70 2.51 0.43 1.03 2.75 0.93	0.20 0.07 0.16 1.10 1.64 0.76 0.70 0.89
D7-V1 D1-V2 D1-V3 D1-V4 D2-V1 D2-V2 D2-V3 D3-V2 D3-V3	6.29 0.04 0.17 0.29 0.42 1.13 1.29 1.46 2.29 2.46	0.26 0.42 1.47 3.29 0.68 0.66 1.37 0.35 0.33	2.18 0.02 0.70 2.51 0.43 1.03 2.75 0.93	0.07 0.16 1.10 1.64 0.76 0.70 0.89
 B D1-V1 D1-V2 D1-V3 D1-V4 D2-V1 D2-V2 D2-V3 D3-V2 D3-V3 	0.04 0.17 0.29 0.42 1.13 1.29 1.46 2.29 2.46	0.42 1.47 3.29 0.68 0.66 1.37 0.35 0.33	0.02 0.70 2.51 0.43 1.03 2.75 0.93	0.16 1.10 1.64 0.76 0.70 0.89
D1-V2 D1-V3 D1-V4 D2-V1 D2-V2 D2-V3 D3-V2 D3-V3	0.17 0.29 0.42 1.13 1.29 1.46 2.29 2.46	1.47 3.29 0.68 0.66 1.37 0.35 0.33	0.70 2.51 0.43 1.03 2.75 0.93	1.10 1.64 0.76 0.70 0.89
D1-V3 D1-V4 D2-V1 D2-V2 D2-V3 D3-V2 D3-V3	0.29 0.42 1.13 1.29 1.46 2.29 2.46	3.29 0.68 0.66 1.37 0.35 0.33	2.51 0.43 1.03 2.75 0.93	1.64 0.76 0.70 0.89
D1-V4 D2-V1 D2-V2 D2-V3 D3-V2 D3-V3	0.42 1.13 1.29 1.46 2.29 2.46	0.68 0.66 1.37 0.35 0.33	0.43 1.03 2.75 0.93	0.76 0.70 0.89
D2-V1 D2-V2 D2-V3 D3-V2 D3-V3	1.13 1.29 1.46 2.29 2.46	0.66 1.37 0.35 0.33	1.03 2.75 0.93	0.70 0.89
D2-V2 D2-V3 D3-V2 D3-V3	1.29 1.46 2.29 2.46	1.37 0.35 0.33	2.75 0.93	0.89
D2-V3 D3-V2 D3-V3	1.46 2.29 2.46	0.35 0.33	0.93	
D3-V2 D3-V3	2.29 2.46	0.33		0.65
D3-V3	2.46		1.98	
				0.62
D5-V1	4 29	0.15	0.68	0.64
		0.00	0.10	0.05
D7-V1	6.29	0.00	1.75	0.09
5 D1-V1	0.04	1.20	0.17	0.42
D1-V2	0.13	1.20	0.74	1.06
D1-V3	0.29	2.12	2.14	2.64
D1-V4	0.46	0.65	0.48	1.87
D2-V1	1.13	0.51	0.80	0.82
D2-V2	1.29	0.57	2.18	0.92
D2-V3	1.46	0.16	0.58	0.64
D3-V1	2.13	0.18	0.52	0.52
D3-V2	2.29	0.17	1.61	0.78
D3-V3	2.46	0.09	0.45	0.64
D5-V1	4.29	0.03	0.50	0.10
D7-V1	6.29	0.00	1.96	0.06
l D1-V1(rai	n) 0.04	0.53	0.07	0.80
D1-V2	0.13	0.58	0.35	1.61
D1-V3	0.29	3.52	1.27	2.98
D1-V4	0.42	0.78	0.36	1.06
D2-V1	1.13	1.16	0.46	1.72
D2-V2	1.29	1.68	1.63	1.33
	1.46	0.50	0.48	0.66
D2-V3	2.13		0.11	0.38
D2-V3 D3-V1			0.46	0.07
	D1-V2 D1-V3 D1-V4 D2-V1 D2-V2 D2-V3	D1-V2 0.13 D1-V3 0.29 D1-V4 0.42 D2-V1 1.13 D2-V2 1.29 D2-V3 1.46 D3-V1 2.13	D1-V20.130.58D1-V30.293.52D1-V40.420.78D2-V11.131.16D2-V21.291.68D2-V31.460.50D3-V12.130.25	D1-V20.130.580.35D1-V30.293.521.27D1-V40.420.780.36D2-V11.131.160.46D2-V21.291.681.63D2-V31.460.500.48D3-V12.130.250.11

0.050 0.431

Table 2.5. The comparison of Haith et.al. model and Aero model of Ethoprop.

(High volatilization group)

Exp	Exp	Exp. #	Time (day)	Observed (%)	Haith (%)	Aero (%)
Week 1	95-1	D1-V1	0.04	- 15 ISI 2017-1		
		D1-V2	0.13			
		D1-V3	0.29		0.52	
		D2-V1	1.13			
		D2-V2	1.29			
		D2-V3	1.46			
		D3-V1	2.13			
		D3-V2	2.29			
		D3-V3	2.46			
		D5-V1	4.29			
		D7-V1	6.29			
Week 4	95-3	D1-V1	0.04			
		D1-V2	0.17			
		D1-V3	0.29			
		D1-V4	0.42			
		D2-V1	1.13			
		D2-V2	1.29		6.96	
		D2-V3	1.46			
		D3-V2	2.29			
		D3-V3	2.46			
		D5-V1	4.29			
		D7-V1	6.29			
Week 7	05.5	DI-VI	0.04			
WOODL /	95-5	D1-V2	0.13			
		D1-V3 D1-V4	0.29			
			0.46			
		D2-V1	1.13			
		D2-V2	1.29			
		D2-V3	1.46			
		D3-V1	2.13		1.32	
		D3-V2	2.29			
		D3-V3	2.46			
		D5-V1	4.29			
		D7-V1	6.29			
Week 12	90-1	D1-V1(rain)				
		D1-V2	0.13			
		D1-V3	0.29			
		D1-V4	0.42			
		D2-V1	1.13			
		D2-V2	1.29			
		D2-V3	1.46			
		D3-V1	2.13			
		D7-V1(rain)				
Week 16	97-1	D1-VI	0.04			
		D1-V2	0.13			
		D1-V3	0.29			
		D2-V1	1.08			
		D2-V2	1.29			
		D2-V3	1.46			
		D3-V1	2.13			1.78
		D3-V2	2.25			
		D3-V3	2.46	0.32		
Week 18	97-2	D1-V1	0.04	3.06		
		D1-V2	0.13	3.43		
		D1-V3	0.29	3.54	2.92	2.94
		D2-V1	1.08	0.82	1.84	1.52
		D2-V2	1.25	0.59	1.81	0.93
		D3-V1	2.13	0.11	1.81	1.44

Table 2.6. The comparison of Haith et.al model and Aero model of Isazofos.

(High volatilization group)

Exp	Exp	Exp. #	Time (day)	Observed (%)	Haith (%)	Aero (%)
Week 1	95-1	D1-V1	(day) 0.04	1.10	0.09	0.31
WOOD I	JJ-1	D1-V2	0.13			0.39
		D1-V2 D1-V3	0.15			0.60
		D2-V1	1.13			1.88
		D2-V1 D2-V2	1.13			1.30
		D2-V2 D2-V3	1.46			0.29
		D3-V1	2.13			0.55
		D3-V1 D3-V2	2.13			0.62
		D3-V2 D3-V3	2.46			0.56
		D5-V3 D5-V1	4.29			0.30
		D3-V1 D7-V1	6.29			0.10
Week 4	95-3	D1-V1	0.29			0.15
WCCK 4	95-5	D1-V1	0.04			1.05
		D1-V2 D1-V3	0.17			1.03
		D1-V3 D1-V4	0.29			0.73
		D1-V4 D2-V1	1.13			
		D2-V1 D2-V2	1.13			
		D2-V2 D2-V3	1.29			
		D2-V3 D3-V2	2.29		2.01	0.68
		D3-V2 D3-V3	2.29			
			4.29			0.06
		D5-V1 D7-V1	6.29			
West 7	95-5		0.29			
Week 7	95-5	D1-V1 D1-V2	0.04			
		D1-V2 D1-V3	0.13			2.53
		D1-V3 D1-V4	0.29			1.81
		D1-V4 D2-V1	1.13			
		D2-V1 D2-V2	1.15			
		D2-V2 D2-V3	1.29			0.66
		D2-V3 D3-V1	2.13			
		D3-V1 D3-V2	2.13			
		D3-V2 D3-V3	2.46			
		D5-V1	4.29			
		D7-V1	6.29			
Week 12	96-1	D1-V1(rain				
WCCK 12	90-1	DI-VI(Iam DI-V2	0.13			
		D1-V2	0.29			
		D1-V3	0.42			
		D1-V4 D2-V1	1.13			
		D2-V1 D2-V2	1.13			
		D2-V2 D2-V3	1.46			
		D2-V3 D3-V1	2.13			
		D3-V1 D7-V1(rain				
		D7-VI(Iam	0.25	0.05	0.40	0.05

0.030 0.337

Exp	Exp	Exp. #	Time	Observed		Aero
			(day)	(%)	(%)	(%)
Week 3	95-2	D1-V1	0.04	0.06	0.00	0.02
		D1-V2	0.13	0.07	0.01	0.03
		D1-V3	0.29	0.06	0.01	0.05
		D2-V1	1.13	0.05	0.00	0.05
		D2-V2	1.25	0.06		0.03
		D2-V3	1.42	0.01	0.00	0.01
		D3-V1	2.08	0.01	0.00	0.01
		D3-V2	2.21	0.01	0.01	0.01
		D5-V1	4.29	0.00	0.01	0.01
		D7-V1	6.29	0.00	0.01	0.00
Week 6	95-4	D1-V1	0.04	0.03	0.00	0.01
		D1-V2	0.13	0.07		0.03
		D1-V3	0.29	0.14	0.02	0.06
		D1-V4	0.46	0.06		0.04
		D2-V1	1.13	0.02	0.00	0.02
		D2-V2	1.29	0.04		0.02
		D2-V3	1.46	0.02		0.01
		D3-V1	2.04	0.00	0.00	0.01
		D3-V2	2.29	0.02	0.01	0.02
		D3-V3	2.46		0.00	0.01
		D5-V1	4.29	0.02	0.02	0.01
		D7-V1	6.29	0.01	0.01	0.00
Week 9	95-6	D1-V1	0.04		0.00	0.01
		D1-V2	0.13	0.02	0.00	0.01
		D1-V3	0.29	0.05	0.00	0.02
		D1-V4	0.46	0.02	0.00	0.03
		D2-V1	1.08		0.00	0.03
Week 13	96-2	D1-V1(rain	0.04	0.01	0.00	0.02
		D1-V2	0.13	0.03	0.00	0.05
		D1-V3(rain	0.29	0.06	0.01	0.11
		D1-V4(rain	0.46	0.03	0.01	0.08
		D2-V1(rain	1.13	0.02	0.00	0.06
		D2-V2(rain	1.29	0.03	0.01	0.07
		D2-V3(rain	1.46	0.02	0.00	0.03
		D3-V1	2.13	0.02	0.00	0.05
		D3-V2	2.29	0.02	0.01	0.05
		D3-V3(rain	2.46	0.01	0.00	0.04
		D5-V1(rain	51 C C C C C C C C C C C C C C C C C C C	0.01	0.01	0.01

0.147 0.246

Table 2.8. The comparison of Haith et.al model and Aero model of Chlorpyrifos.

(Low volatilization group)

Exp	Exp	Exp. #	Time	Observed		Aero
			(day)	(%)	(%)	(%)
Week 1	95-1	D1-V1	0.04	0.58	0.06	0.21
		D1-V2	0.13	0.73	0,10	0.28
		D1-V3	0.29		0.13	0.43
		D2-V1	1.13	3.15	0.55	1.50
		D2-V2	1.29	4.12	1.19	
		D2-V3	1.46	0.37		
		D3-V1	2.13	0.67	0.89	0.50
		D3-V2	2.29			
		D3-V3	2.46	0.61	0.52	0.52
		D5-V1	4.29	0.54	1.80	0.28
		D7-V1	6.29	0.43	1.70	0.15
Week 4	95-3	D1-V1	0.04	0.26	0.02	0.10
		D1-V2	0.17	0.95	0.55	0.74
		D1-V3	0.29	2.25	1.98	1.14
		D1-V4	0.42	0.48	0.34	0.53
		D2-V1	1.13	0.58	0.82	0.57
		D2-V2	1.29	1.51	2.18	0.74
		D2-V3	1.46	0.29	0.74	0.56
		D3-V2	2.29	0.39	1.57	0.62
		D3-V3	2.46	0.16	0.54	0.66
		D5-V1	4.29	0	0.08	0.07
		D7-V1	6.29	0	1.37	0.18
Week 7	95-5	D1-V1	0.04		0.14	0.28
		D1-V2	0.13	1.21	0.58	0.71
		D1-V3	0.29	1.66	1.68	1.83
		D1-V4	0.46			
		D2-V1	1.13			0.66
		D2-V2	1.29	0.84	1.73	0.77
		D2-V3	1.46		0.45	
		D3-V1	2.13			0.50
		D3-V2	2.29			
		D3-V3	2.46			
		D5-V1	4.29			
		D7-V1	6.29			
Week 12	96-1	D1-V1(rain				
		D1-V2	0.13		0.25	
		D1-V3	0.29			
		D1-V4	0.42			
		D2-V1	1.13			
		D2-V2	1.29			
		D2-V2	1.46			
		D3-V1	2.13			
		D7-V1(rain				
		2. Contraint		-15		0.755202

0.100 0.355

 \mathbb{R}^2

Table 2.9. The comparison of Haith et.al model and Aero model of Isofenphos.

(Low volatilization group)

]	Exp	Exp	Exp. #	Time (day)	Observed (%)	Haith (%)	Aero (%)
1	Week 4	95-3	D1-VI	0.04	0.00	0.00	0.04
			D1-V2	0.17	0.00	0.09	0.25
			D1-V3	0.29	0.00	0.34	0.36
			D1-V4	0.42	0.00	0.05	0.16
			D2-V1	1.13	0.24	0.14	0.13
			D2-V2	1.29	0.00	0.40	0.16
			D2-V3	1.46	0.00	0.13	0.11
			D3-V2	2.29			
			D3-V3	2.46			
			D5-V1	4.29			
			D7-V1	6.29			
9	Week 7	95-5	D1-V1	0.04			
	HOUL /	100	D1-V2	0.13			
			D1-V2	0.29			
			D1-V4	0.46			
			D2-V1	1.13			
			D2-V1 D2-V2	1.13			
			D2-V3	1.46			
			D3-V1	2.13			
			D3-V2	2.29			
			D3-V3	2.46			
			D5-V1	4.29			
	aan e waa	0.000	D7-V1	6.29			
	Week 12	96-1	D1-V1(rain	5			
			D1-V2	0.13			
			D1-V3	0.29			
			D1-V4	0.42			
			D2-V1	1.13	0.20	0.05	0.32
			D2-V2	1.29	0.32	0.20	0.24
			D2-V3	1.46	0.09	0.06	0.12
			D3-V1	2.13	0.02	0.01	0.06
			D7-V1(rain	6.29	0.00	0.06	0.00
	Week 16	97-1	D1-V1	0.04	0.36	0.03	0.27
			D1-V2	0.13	0.74	0.09	0.48
			D1-V3	0.29		0.17	0.58
			D2-V1	1.08	0.21	0.06	0.32
			D2-V2	1.29		0.20	0.33
			D2-V3	1.46		0.07	0.13
			D3-V1	2.13			
			D3-V2	2.25			
			D3-V3	2.46			
	Week 18	97-2	D1-V1	0.04			
	WOOL ID	21-4	D1-V2	0.13			
			D1-V3	0.29			
			D2-V1	1.08			
				1.25			
			D2-V2	2.13			
			D3-V1	2.13			
		07.2	D3-V2				
	Week 20	97-3	D1-V1	0.08			
			D1-V2	0.13			
			D1-V3	0.29			
			D2-V1	1.08			
			D2-V2	1.29			
			D2-V3	1.42			
			D3-V1	2.08			
			D3-V2	2.29	0.13	0.12	2 0.07

R²

0.005 0.227

Table 2.10. The comparison of Haith et al. model and Aero model of Trichlorfon.

(Low volatilization group)

Exp	Exp	Exp. #	Time	Observed		Aero
			(day)	(%)	(%)	(%)
Week 3	95-2	D1-V1	0.04	0.27	0.055	0.05
		D1-V2	.0.13	0.37	0.154	0.08
		D1-V3	0.29			0.12
		D2-V1	1.13	0.08		0.12
		D2-V2	1.25			0.09
		D2-V3	1.42		0.006	0.02
		D3-V1	2.08		0.031	0.03
		D3-V2	2.21	0.02		0.03
		D5-V1	4.29		0.142	0.03
		D7-V1	6.29		0.111	0.01
Week 6	95-4	D1-V1	0.04			0.03
		D1-V2	0.13			0.07
		D1-V3	0.29			0.15
		D1-V4	0.46		0.082	0.10
		D2-V1	1.13			0.06
		D2-V2	1.29		0.169	0.06
		D2-V3	1.46			0.03
		D3-V1	2.04			0.02
		D3-V2	2.29			
		D3-V3	2.46	0.07		
		D5-V1	4.29	0.08	0.171	0.03
		D7-V1	6.29	0.06	0.119	0.01
Week 9	95-6	D1-V1	0.04	0.02	0.002	0.02
		D1-V2	0.13	0.11	0.005	
		D1-V3	0.29	0.17	0.014	0.05
		D1-V4	0.46	0.07	0.018	0.06
		D2-V1	1.08	0.04	0.070	0.08
Week 13	96-2	D1-V1(rain	i) 0.04	0.03	0.012	0.05
		D1-V2	0.13	0.1	0.061	0.12
		D1-V3(rain	i) 0.29	0.08	0.201	0.25
		D1-V4(rain	i) 0.46	0.04	0.101	0.17
		D2-V1(rai	i) 1.13	0.04	0.062	0.15
		D2-V2(rain	ij 1.29	0.05	0.179	0.18
		D2-V3(rai	i) 1.46	0.02	0.058	0.09
		D3-V1	2.13	0.08	0.025	0.14
		D3-V2	2.29	0.05	0.146	0.14
		D3-V3(rai	i) 2.46	0.02	0.046	0.11
		D5-V1(rain			0.108	0.05
D ²					0.0738	0.0181

 R^2

0.0738 0.0181

Pesticide	Exp	Observed (%)	Haith (%)	Aero (%)
Bendiocarb	95-2	1.53	1.83	1.15
	95-4	2.97	2.52	1.31
	95-6	0.59	0.28	0.43
	96-2	1.41	2.33	3.02
	R ²		0.65	0.03
Diazinon	95-1	17.02	11.69	6.54
	95-3	8.72	12.89	7.29
	95-5	6.88	12.16	10.47
	96-1	9.22	5.19	10.63
	R ²		0.01	0.51
Ethoprop	95-1	22.17	29.67	7.47
	95-3	14.25	32.49	9.39
	95-5	9.99	31.17	13.78
	96-1	19.09	14.90	15.02
	97-1	16.77	16.24	17.38
	97-2	11.64	15.11	12.94
	97-3	12.12	16.86	7.46
	R ²		0.00	0.01
Isazofos	95-1	20.62	12.01	6.74
	95-3	5.44	13.16	7.34
	95-5	6.62	12.52	10.48
	96-1	8.58	5.44	10.45
	R ²		0.00	0.31

Table 2.11. Comparison of Haith *et al.* model with Aero model in the sum of each set of experiment.

Table 2.11 (C	Continued)
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Pesticide	Exp	Observed (%)	Haith (%)	Aero (%)
Carbaryl	95-2	0.33	0.08	0.23
	95-4	0.44	0.11	0.26
	95-6	0.09	0.00	0.10
	96-2	0.26	0.08	0.59
	R ²		0.93	0.05
Chlorpyrifos	95-1	13.67	9.10	5.67
	95-3	6.87	10.18	5.91
	95-5	6.46	9.51	8.33
	96-1	5.98	3.87	8.00
	R ²		0.09	0.47
Isofenphos	95-3	0.24	1.80	1.41
	95-5	0.79	1.62	2.04
	96-1	1.22	0.59	2.17
	97-1	2.69	0.67	2.60
	97-2	2.21	0.59	1.87
	97-3	2.04	0.79	1.10
	R ²		0.69	0.11
Trichlorfon	95-2	1.19	0.89	0.56
	95-4	1.05	1.22	0.63
	95-6	0.41	0.11	0.24
	96-2	0.57	1.00	1.45
	R ²		0.45	0.01

Pesticide	Haith	Aero	Haith	Aero
	R ² ,	R ² ,	R ² , sum	R ² , sum
	Individually	Individually		
*Bendiocarb	0.35	0.01	0.65	0.03
*Diazinon	0.05	0.43	0.01	0.51
*Ethoprop	0.11	0.54	0.00	0.01
*Isazofos	0.03	0.34	0.00	0.31
Carbaryl	0.15	0.25	0.93	0.05
Chlorpyrifos	0.10	0.36	0.09	0.47
Isofenphos	0.01	0.23	0.69	0.11
Trichlorfon	0.07	0.02	0.45	0.01

Table 2.12. R² values of both models (Haith & Aero).

* Classified as high volatilization group in Haith et al., 2002. The others are low volatilization group.

	Half Life from	Half Life from	Half Life in	Field	
	a (day)	Haith et al.	soil from	Dissipation	
		(day)	USDA (day)	Half Life from	
				USDA (day)	
Bendiocarb	2.12	2.9	0.94~3.5	3~21	
Diazinon	1.05	39.0	39	2.8~48	
Ethoprop	0.73	24.0	24	4~87	
Isazofos	1.14	40.5	45	2.5~48.4	
Carbaryl	1.16	17.0	7~27	4~13	
Chlorpyrifos	1.44	30.5	30.5	1.3~139	
Isofenphos	1.20	93.0	59~127	12~365	
Trichlorfon	1.28	6.4	1.8~6.4	< 1~2.2	

Table 2.13. Comparison of degradation (Half Life) from different references.

* Half life data of USDA is from Agricultural Research Service. 2001.

	Half Life of	Half Life of	Shortest Half
	Hydrolysis	Photolysis	Life of
	from USDA	from USDA	(Hydrolysis +
	(day)	(day)	Photolysis)
			Calculated.
			from USDA
			(day)
Bendiocarb	0.03~5.0	0.29~37.3	0.03
Diazinon	11.8~77.0	4.6~5.1	3.30
Ethoprop	19.8	23.9	10.83
Isazofos	14.0~115.0	4.0~40.8	3.11
Carbaryl	10.5	45	8.51
Chlorpyrifos	15.8~77.0		15.75
Isofenphos	30.1	72.2~99.0	21.26
Trichlorfon	34.7~103.4	11.8~110.2	0.02

Table. 2.13. (Continued)

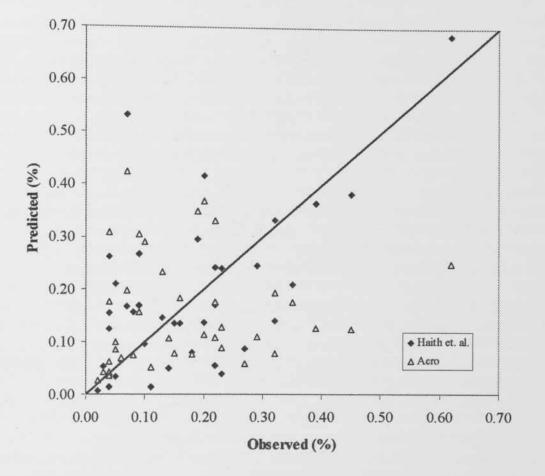


Figure 2.9. The comparison of predicted pesticide volatilization with observed volatilization of Bendiocarb.

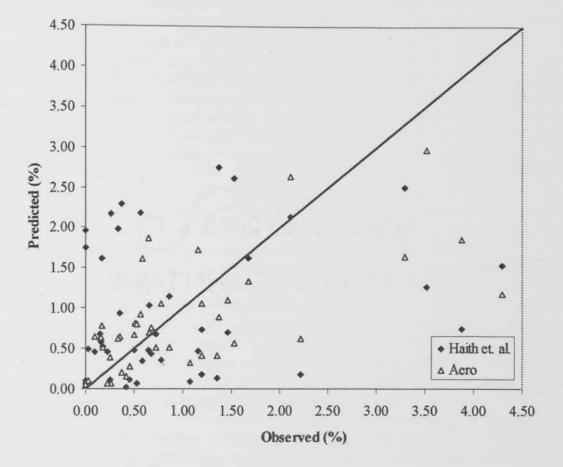


Figure 2.10. The comparison of predicted pesticide volatilization with observed volatilization of Diazinon.

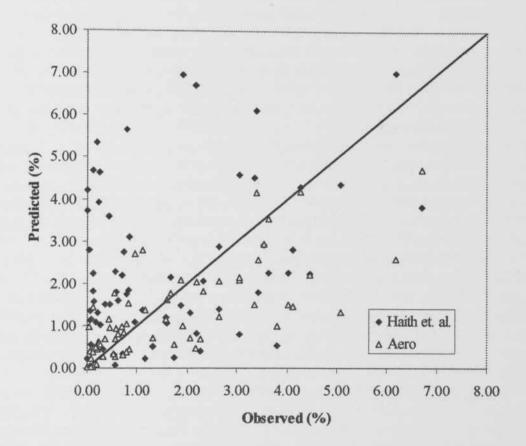


Figure 2.11. The comparison of predicted pesticide volatilization with observed volatilization of Ethoprop.

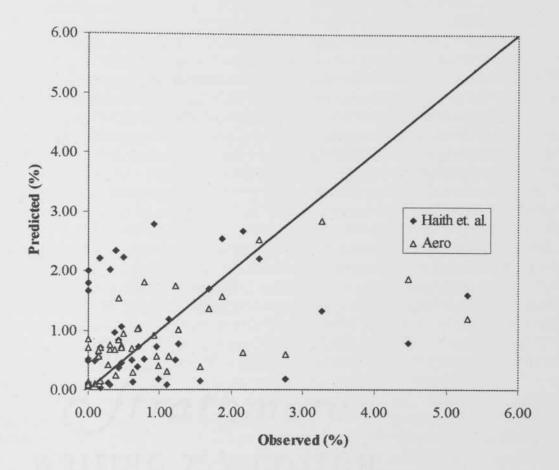


Figure 2.12. The comparison of predicted pesticide volatilization with observed volatilization of Isazofos.

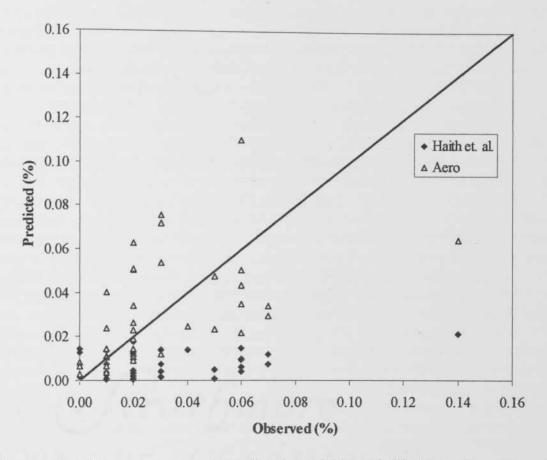


Figure 2.13. The comparison of predicted pesticide volatilization with observed volatilization of Carbaryl.

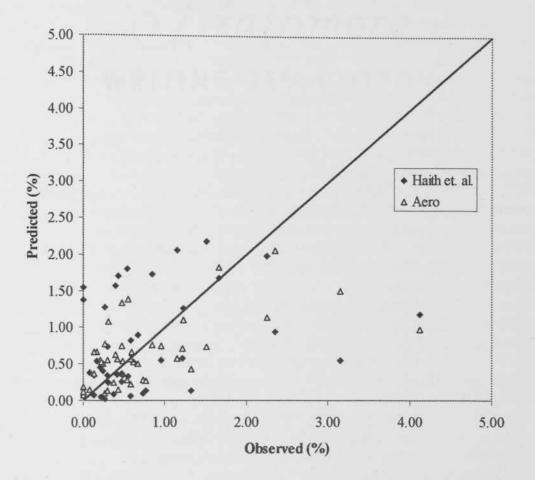


Figure 2.14. The comparison of predicted pesticide volatilization with observed volatilization of Chlorpyrifos.

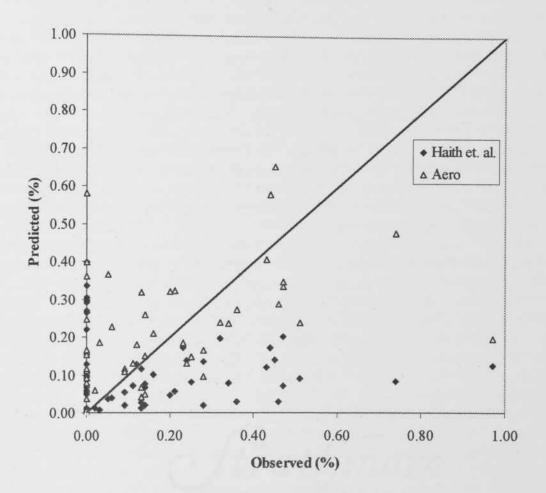


Figure 2.15. The comparison of predicted pesticide volatilization with observed volatilization of Isofenphos.

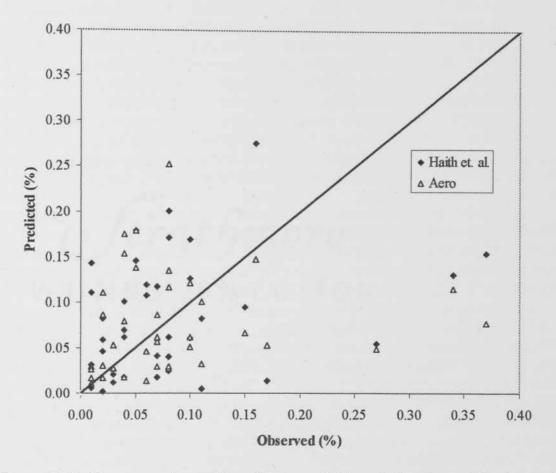


Figure 2.16. The comparison of predicted pesticide volatilization with observed volatilization of Trichlorfon.

Symbols listed in this chapter

- a = degradation rate of pesticide, day-1
- $b = adsorption parameter, m^3 m^{-2}$
- $C_p =$ solute concentration, mole/m³
- C_s = concentration of pesticide on the solid phase, mole g⁻¹
- d = vapor concentration of pesticide, mole/m³
- d_1 = vapor concentration of pesticide at z_1 , mole/m³
- d_2 = vapor concentration of pesticide at z_2 , mole/m³
- d_s = saturated pesticide vapor pressure, the same unit as e_s, kPa or mole m⁻³ $d_{s,T_{a_o}}$ = saturated pesticide vapor pressure at temperature T_{a_o} , kPa

 $F_{w,H}$ · Δt = water evapotranspiration during time Δt , mm

- $e_1 =$ vapor concentration of water at z_1 , mole m⁻³
- $e_2 =$ vapor concentration of water at z_2 , (mole m⁻³)
- e's = saturated water vapor pressure at air temperature T, kPa
- F = vertical flux of vapor, mole m⁻² sec⁻¹
- F_p = vapor flux of pesticide, mole m⁻² hr⁻¹
- F_w = vapor flux of water, mole m⁻² hr⁻¹
- h = Henry's law constant, dimensionless
- K = the adsorption partition coefficient, m³ g⁻¹
- $k = volatilization constant, mm^{-1}$
- K_f = constant, dimensionless, 1.06 (Haith, 2002)
- k_z = mean eddy diffusivity coefficient or transport coefficient, m² sec⁻¹
- m = constant, 0.8 (Haith, 2002)
- $p = vapor concentration, mole m^{-3}$
- P_A = pesticide on plant leaves per unit area, mole m⁻²

 $(P_A)_o$ = the initial mass of pesticide per unit area at time t = 0, mole m⁻²

 $(P_A)_{\bar{t}}$ = the amount of pesticide remaining on the leaves at time \bar{t} , mole m⁻²

 $(P_{A,H})_t$ = pesticide available for volatilization on vegetation at the beginning time step t, g ha⁻¹

 $(P_{A,H})_{t+1}$ = pesticide available for volatilization on vegetation at the beginning of time step t+1, g ha⁻¹

R = gas constant, 8.314 J mole⁻¹ K⁻¹

 $RH = relative humidity during time \Delta t$, dimensionless

S = weight of sorbent in contact with the pesticide, g m⁻²

T = air temperature during time Δt , °C

 T_a = air temperature during time Δt , K

 T_{a_0} = temperature at the addressed $d_{s,T_{a_0}}$, K

 T_t = temperature during time Δt , °C

t' = time period between each sampling period, hr

t = time since initial pesticide spraying, day

W = amount of water per unit area on the plant leaves, m^3/m^2

z = z coordinate in the vertical direction, m

 z_1 = vertical direction at height 1, m

 z_2 = vertical direction at height 2, m

 $\Delta(P_{A,v})_{\tilde{t}}$ = pesticide loss (volatilization) at time \tilde{t} in short period of time Δt of Aero model, mole m⁻²

 $\Delta(P_{A,H})_i$ = pesticide vaporized from surface vegetation during time Δt at time step t of Haith model, g ha⁻¹

 ΔZ_b = compressibility factor at boiling point, dimensionless, 0.97 (Grain, 1982)

 $\Delta t =$ sampling period of pesticide volatilization, day

 α = degradation rate of pesticide on vegetation surface, hr⁻¹

 $\lambda_{p,t}$ = latent heat of vaporization of the chemical during time Δt at time step t, J g^-1 or kJ kg^{-1}

 $\mathcal{\lambda}_{{}_{p,T_{a_0}}}$ = latent heat of pesticide vaporization at temperature T_{a_0} , J/g

 $\lambda_{w,t}$ = latent heat of vaporization of water during time Δt at time step t, J g^{-1} or kJ kg^{-1}

	Append	ix 2.1	. Input	Data
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Experime	and the second se	A CONTRACTOR OF A CONTRACTOR O	une 14 - 20, 199	1		
Period	Air Temperature , T _b °C	Duration, hr	Penman Evaporation During Duration, $F_{w,H} \Delta t$, mm	Lowest temperature of a day, Td _b °C	Solar Radiation, Rs, KJ m ⁻² hr ⁻¹	Wind speed, u, ms ⁻¹
D1-V1	23.1	1	0.17	13.3	990	1.7
D1-V2	24.1	2	0.25	13.3	836	1.1
D1-V3	22.8	4	0.35	13.3	631	2
D2-V1	20	4	1.70	7.7	2029	2.8
D2-V2	28.9	4	2.60	7.7	2520	2.4
D2-V3	19.2	4	0.28	7.7	590	1.3
D3-V1	29.8	4	1.94	6.6	2033	1
D3-V2	36.5	4	3.66	6.6	3216	2.1
D3-V3	24.2	4	1.47	6.6	1638	1.9
D5-V1	36.9	4	3.41	12.2	2908	2.8
D7-V1	38.3	4	3.31	18.3	3022	1.8

Experiment Set	95-2 week	3 July 6 - 1	2, 1995			
Period	Air Temperature, T _b °C	Duration, hr	Penman Evaporation During Duration, $F_{w,H}\Delta t$, mm	Lowest temperature of a day, Td _b °C	Solar Radiation, Rs, KJ m ⁻² hr ⁻¹	Windspeed, u, ms ⁻¹
D1-V1	34.5	1	0.70	19.4	2644	2.6
D1-V2	39.3	2	1.69	19.4	2892	2.9
D1-V3	34	4	1.74	19.4	1622	3
D2-V1	28	4	1.14	20.5	1456	1.9
D2-V2	33.6	3	1.78	20.5	2286	2.8
D2-V3	26.1	3	0.13	20.5	442	1.2
D3-V1	29.9	3	0.60	19.4	1040	1.6
D3-V2	36.3	2	1.26	19.4	2325	2.3
D5-V1	35	4	2.83	12.7	2539	2.4
D7-V1	35.8	4	2.66	16.1	2541	1.5

Appendix 2.1. (Continued)

Experiment Set	95-3 week	4 July 13 -	19, 1995			
Period	Air Temperature , T _b °C	Duration, hr	Penman Evaporation During Duration, $F_{w,H}\Delta t$, mm	Lowest temperature of a day, Td _b °C	Solar Radiation, Rs, KJ m ⁻² hr ⁻¹	Windspeed, u, ms ⁻¹
D1-V1	23.1	1	0.05	17.7	309	1.9
D1-V2	30.7	3	1.09	17.7	1529	2.3
D1-V3	37.6	4	3.17	17.7	2805	2.5
D1-V4	29.2	3	0.73	17.7	1137	2.1
D2-V1	34.3	4	1.50	19.4	1575	1.5
D2-V2	40.4	4	3.42	19.4	2964	2.3
D2-V3	33.1	4	1.48	19.4	1483	2.4
D3-V2	37.6	4	2.81	23.3	2669	2
D3-V3	30.1	4	1.25	23.3	1485	2.9
D5-V1	25.3	4	0.23	17.2	476	1.1
D7-V1	33.7	4	3.09	15.5	2835	2.8

Appendix 2.1. (Continued)

Append	lix 2.1.	(Continued)	

Experiment Set	95-4 week	6 August 1	3 - 19, 1995			
Period	Air Temperature, T _b °C	Duration, hr	Penman Evaporation During Duration, $F_{w,H}\Delta t$, mm	Lowest temperature of a day, Td _b °C	Solar Radiation, Rs, KJ m ⁻² hr ⁻¹	Windspeed, u, ms ⁻¹
D1-V1	30.2	1	0.31	15.5	1425	1.1
D1-V2	35.2	2	1.20	15.5	2299	1.6
D1-V3	37.1	4	3.28	15.5	2924	2.3
D1-V4	29.8	4	1.32	15.5	1344	2.6
D2-V1	29.3	4	1.03	13.3	1193	1.3
D2-V2	37.5	4	2.23	13.3	2031	1.6
D2-V3	31.2	4	0.67	13.3	768	1.5
D3-V1	29.7	2	0.34	20	929	1.7
D3-V2	35.5	4	1.98	20	1872	2.4
D3-V3	31.1	4	0.79	20	940	2.2
D5-V1	38.3	4	3.05	22.2	2775	2.4
D7-V1	33.6	4	3.11	12.2	2874	2.3

Experiment Set	95-5 week 7 August 22 - 28, 1995							
Period	Air Temperature, T _t , °C	Duration, hr	Penman Evaporation During Duration, $F_{w,H}\Delta t$, mm	Lowest temperature of a day, Td _t , °C	Solar Radiation, Rs, KJ m ⁻² hr ⁻¹	Windspeed, u, ms ⁻¹		
D1-V1	27.7	1	0.30	16.6	1321	2.8		
D1-V2	31.1	2	1.13	16.6	2101	3.6		
D1-V3	31.7	4	3.24	16.6	2838	5		
D1-V4	23.8	4	0.98	16.6	1237	3.7		
D2-V1	27.6	4	1.45	8.8	1603	1.3		
D2-V2	35.3	4	3.12	8.8	2753	2.3		
D2-V3	25.8	4	1.16	8.8	1251	2.2		
D3-V1	26.3	4	1.04	15	1251	2.2		
D3-V2	31.1	4	2.77	15	2406	4.7		
D3-V3	23.2	4	1.04	15	1253	3.9		
D5-V1	27	4	1.03	5.5	1127	1.5		
D7-V1	36.7	4	3.15	8.3	2824	1.7		

Appendix 2.1. (Continued)

Appendix 2.1. (Continued)

Experiment Set	95-6 week 9 September 19 - 20, 1995								
Period	Air Temperature, T _t , °C	Duration, hr	Penman Evaporation During Duration, $F_{w,H}\Delta t$, mm	Lowest temperature of a day, Td _b , °C	Solar Radiation, Rs, KJ m ⁻² hr ⁻¹	Windspeed, u, ms ⁻¹			
D1-V1	15	1	0.07	3.3	46	1.8			
D1-V2	15.6	2	0.16	3.3	103	1.8			
D1-V3	18.5	4	0.37	3.3	293	1.7			
D1-V4	18.9	4	0.48	3.3	199	2.6			
D2-V1	26	3	1.41	4.4	1716	3.5			

Appendix 2.1. (Continued)

Experiment Set	96-1 week 12 May 14 - 20, 1996								
Period	Air Temperature, T_b °C	Duration, hr	$\begin{array}{c} \text{Penman} \\ \text{Evaporation} \\ \text{During} \\ \text{Duration,} \\ F_{w,H} \Delta t, \\ \text{mm} \end{array}$	Lowest temperature of a day, Td _b °C	Solar Radiation, Rs, KJ m ⁻² hr ⁻¹	Windspeed, u, ms ⁻¹			
D1-V1(rain)	9.6	1	0.23	-2.8	1613	1.2			
D1-V2	16.1	2	0.91	-2.8	2370	1.5			
D1-V3	21.2	4	2.75	-2.8	2879	3			
D1-V4	18.7	3	0.87	-2.8	1348	2.1			
D2-V1	14.4	4	1.35	-1.2	1934	1.2			
D2-V2	26.8	4	3.00	-1.2	3055	1.6			
D2-V3	21.2	4	1.10	-1.2	1222	2			
D3-V1	12.3	4	0.37	1.1	735	1.7			
D7-V1(rain)	25	4	1.00	15.5	1279	2.2			

Appendix 2.1. (Continued)

Experiment Set	96-2 week 13 May 22 - 28, 1996								
Period	Air Temperature, T _b °C	Duration, hr	Penman Evaporation During Duration, $F_{w,H}\Delta t$, mm	Lowest temperature of a day, Td _b °C	Solar Radiation, Rs, KJ m ⁻² hr ⁻¹	Windspeed, u, ms ⁻¹			
D1-V1(rain)	19.9	1	0.29	8.8	1629	1.2			
D1-V2	25	2	1.17	8.8	2529	2.1			
D1-V3(rain)	28.1	4	3.38	8.8	3221	3.4			
D1-V4(rain)	25.2	4	1.97	8.8	2015	3			
D2-V1(rain)	20.8	4	1.62	7.2	1996	1.7			
D2-V2(rain)	27.1	4	3.52	7.2	3297	4.1			
D2-V3(rain)	23.7	4	1.37	7.2	1498	2.4			
D3-V1	19.2	4	0.78	15	1300	2.5			
D3-V2	27.5	4	3.16	15	3227	3.2			
D3-V3(rain)	22	4	1.32	15	1672	3.4			
D5-V1(rain)	25.5	4	3.19	2.2	3181	2.7			
D7-V1(rain)	25.1	4	2.65	9.4	2822	2.4			

Appendix 2.1. (Continued)

Experiment Set	97-1 week 16 May 28 - May 30, 1997								
Period	Air Temperatur e, T _t , °C	Duration, hr	Penman Evaporation During Duration, $F_{w,H}\Delta t$, mm	Lowest temperature of a day, Td _b °C	Solar Radiation, Rs, KJ m ⁻² hr ⁻¹	Windspeed, u, ms ⁻¹			
D1-V1	20.4	1	0.59	9	2945	1.3			
D1-V2	23.3	2	1.50	9	3381	1.8			
D1-V3	26.2	4	2.68	9	2892	1.5			
D2-V1	19.9	3	1.21	11	2158	1.3			
D2-V2	26.7	4	3.08	11	3263	1.9			
D2-V3	25.3	4	1.18	11	1279	2.7			
D3-V1	16.1	4	0.32	15	936	1.8			
D3-V2	18.9	3	0.45	15	1129	2			
D3-V3	21.2	4	0.40	15	754	2.7			

Appendix 2.1. (Continued)

Experiment Set	97-2 week 18 July 8 - 10, 1997								
Period	Air Temperature, T _b °C	Duration, hr	Penman Evaporation During Duration, $F_{w,H}\Delta t$, mm	Lowest temperature of a day, Td _b °C	Solar Radiation, Rs, KJ m ⁻² hr ⁻¹	Windspeed, u, ms ⁻¹			
D1-V1	22.8	1	0.53	15	2620	1.7			
D1-V2	26.3	2	1.08	15	2499	1.3			
D1-V3	28	4	1.72	15	1936	1.5			
D2-V1	26.5	3	1.20	18	1868	2.5			
D2-V2	28.7	3	1.13	18	1659	2.5			
D3-V1(rain)	21.4	4	1.48	16	1982	2.6			
D3-V2	24.9	4	2.10	16	2453	2.2			
D3-V3	25.4	4	1.04	16	1368	1.6			

Appendix 2.1. (Continued)

Experiment Set	97-3 week 20 August 26 - 28, 1997								
Period	Air Temperature, T _b °C	Duration, hr	$\begin{array}{c} \text{Penman} \\ \text{Evaporation} \\ \text{During} \\ \text{Duration,} \\ F_{w,H} \Delta t, \\ mm \end{array}$	Lowest temperature of a day, Td _b °C	Solar Radiation, Rs, KJ m ⁻² hr ⁻¹	Windspeed, u, ms ⁻¹			
D1-V1	34.3	2	1.35	12	2624	1.2			
D1-V2	36.4	2	1.35	12	2546	1.2			
D1-V3	30.7	4	1.27	12	1383	1.2			
D2-V1(rain)	27.8	3	0.97	17	1551	1.6			
D2-V2	31.7	4	2.10	17	2082	2.4			
D2-V3	27.4	3	0.71	17	1094	2.9			
D3-V1	25.7	3	0.42	17	911	1.3			
D3-V2	30.9	4	1.46	17	1551	2			

Pesticide	Experiment	Application (g ha ⁻¹)
Bendiocarb	95-2	5832
	95-4	4666
	95-6	4666
	96-2	5832
Diazinon	95-1	4204
	95-3	4605
	95-5	5146
	96-1	5676
Ethoprop	95-1	6327
•	95-3	6907
	95-5	7719
	96-1	8514
	97-1	10256
	97-2	7137
	97-3	6883
Isazofos	95-1	2099
	95-3	2302
	95-5	2573
	96-1	2838

Appendix 2.2. Initial amount of pesticide applied in each set of experiment

Appendix 2.2. (Continued)

Pesticide	Experiment	Application (g ha ⁻¹)
Carbaryl	95-2	2803
	95-4	2243
	95-6	2243
	96-2	2803
Chlorpyrifos	95-1	6997
	95-3	7674
	95-5	8577
	96-1	9460
Isofenphos	95-3	2301
	95-5	2573
	96-1	2838
	97-1	3419
	97-2	2379
	97-3	2294
Trichlorfon	95-2	11484
	95-4	9187
	95-6	9187
	96-2	11484

Appendix 2.3. Penman equation (Haith, 2002; Jensen ,1990)

The Penman equation provides an estimate of water evapotranspiration for "well water short grass" (Jensen, 1990). The Penman equation could be described as:

$$F_{w,H,t} = \left(\frac{\Delta_t}{\Delta_t + \gamma_t} \cdot (Rn_t - G_t) + \frac{\gamma_t}{\Delta_t + \gamma_t} \cdot 268 \cdot (1 + 0.53 \cdot u_t) \cdot (e_{s,t}' - e_{2,t}')\right) / \lambda_{w,t} \quad (A2.1)$$

 $F_{w,H,t} =$ evapotranspiration during hour t, mm $\lambda_{w,t} =$ latent heats of vaporization water during hour t, J g⁻¹. $\Delta_t =$ slope of saturation vapor pressure curve during hour t, kPa °C⁻¹ $\gamma_t =$ psychrometric constant during hour t, kPa °C⁻¹ $Rn_t =$ net radiant energy available at surface during hour t, kJ m⁻² h⁻¹ $G_t =$ net sensible heat flux from the surface to soil during hour t, kJ m⁻² h⁻¹ $u_t =$ mean wind velocity during hour t, m s⁻¹ $e'_{s,t} =$ saturated water vapor pressure during hour t, kPa

Parameters in Equation A2.1 are given in the following equations as described by Jensen, 1990 and Haith, 2002.

$\lambda_{\rm wt} = 2501 - 2.361 {\rm T_t}$	(A2.2)	
$e'_{s} = 3.38639[(0.00738 \text{ T} + 0.8072)^{8} - 0.000019 1.8 \text{ T} + 48 + 0.001316]$	(A2.3)	
$\Delta_{\rm t} = 0.2(0.00738 {\rm T_t} + 0.8072)^7 - 0.000116$	(A2.4)	
$\gamma_t = 1.013 \text{ P} / (0.622 \lambda_{w,t})$	(A2.5)	
P = 101.3 - 0.01055 EL	(A2.6)	

where

 $T_t = air temperature during hour t, °C$

P = mean atmospheric pressure at the site, kPa

EL = elevation, m

The net radiant energy term (Rn_t - G_t) in Equation A2.1 is given by incoming solar radiation minus reflected radiation and net thermal radiation. Only the first of these is generally known, and net energy is usually estimated from regression equation (Haith, 2002). Jensen (1990) provide a number of these equations determined for various covers and locations, including the following that was obtained for grass in Minnesota:

$$Rn_t = 0.79 Rs_t - 470$$
 (A2.7)

where

 $Rs_t = solar radiation during hour t (kJ m⁻² h⁻¹)$

Appendix 2.4. Ideal Gas Law

PV = nRT

ог

$$p = \frac{n}{V}RT$$

where

P = gas pressure, Pa

V = volume of air, m^3

n = mole of gas, mole

R = gas constant, 8.314 J mole⁻¹ K⁻¹

T = temperature, K

References

Agricultural Research Service. 2001. Pesticide properties database. http://wizard.arsusda.gov/acsl/acslhome.html.

Caro, J.H., and A.W. Taylor. 1971. Pathways of loss of dieldrin from soils under field conditions. J. Agric. Food Chem. 19:379-384.

Clark, J. M., Roy, G. R., Doherty, J. J., Curtis, A. S., Cooper, R. J. 2000. Hazard evaluation and management of volatile and dislodgeable foliar pesticide residues following application to turfgrass. In: Clark, J. M, Kenna, M. P. (ed.). *Fate and Management of Turfgrass Chemicals*. ACS Symposium Series 743. American Chemical Society, Washington, DC. pp. 294-312.

Cliath, M.M., and Spencer, W.F. 1971. Movement and persistence of dieldrin and lindane in soil as influenced by placement and irrigation. *Soil Sci. Soc. Am. J.* 35:791-795.

Cooper, R. J., Jenkins, J. J., Curtis, A. S. 1990. Pendimethalin volatility following application to turfgrass. *Journal of Environmental Quality* 19(3):508-513.

Dingman, S.L. 2002. *Physical Hydrology*. Prentice-Hall, Inc. New Jersey. pp. 272 to 285.

Eagleson, P.S. 1970. Dynamic Hydrology. McGraw-Hill Book Company, New York.

Farmer, W.J., K. Igue, W.F. Spencer, and Martin, J.P. 1972. Volatility of organochlorine insecticides from soil: I. Effect of concentration, temperature, air flow rate, and vapor pressure. *Soil Sci. Soc. Am. J.* 36:443-447.

Farmer, W.J., W.F. Spencer, R.A. Shepherd, and Cliath, M.M. 1974. Effect of flooding and organic matter application on DDT residues in soil. *J. Environ. Qual.* 3:343-346.

Harper, L.A., A.W. white, Jr., R.R. Bruce, A.W. Thomas, and Leonard, R.A. 1976. Soil and microclimate effects on trifluralin volatilization. *J. Environ. Qual.* 5:236-242.

Haith, D.A., Lee, P.-C., Clark, J. M., Roy, G.R., Imboden, M. J. and Walden, R. R. 2002. Modeling pesticide volatilization from turf. *Journal of Environmental Quality* 31(3):724-729.

Jensen, M. E., Burman, R. D., Allen, R. G. (ed.) 1990. Evapotranspiration and irrigation water requirements. ASCE Manual No. 70. American Society of Civil Engineers, New York, NY.

Jury, W.A. Spencer, W.F., and Farmer, W.J. 1983. Behavior assessment model for trace organics in soil: I. Model description. *J.Environ. Qual.*, Vol 12, No. 4: 558-564.

Monteith, J. L. 1973. Principles of Environmental Physics. American Elsevier Pub. New York, NY. Murphy, K. C., Cooper, R. J., Clark, J. M. 1996a. Volatile and dislodgeable residues following trichlorfon and isazofos application to turfgrass and implications for human exposure. *Crop Science* 36(6): 1446-1454.

Murphy, K. C., Cooper, R. J., Clark, J. M. 1996b. Volatile and dislodgeable residues following triadimeton and MCPP application to turfgrass and implications for human exposure. *Crop Science* 36(6): 1455-1461.

Prueger, J. H., Hatfield, J. L., Sauger, T. J. 1999. Field-scale metolachlor volatilization flux estimates from broadcast and banded application methods in central Iowa. *Journal of Environmental Quality* 28(1): 75-81.

Spencer, W.F., and Farmer, W.J. 1980. Assessment of the vapor behavior of toxic organic chemical. P. 142-162. In R. Hague (ed.) Dynamics, exposure, and hazardous assessments of toxic chemicals in the environment. Ann Arbor Sci., Ann Arbor. MI.

Timmons, M.B. and R.S. Gates. 1986. Economic Optimization of Boiler Production. TRANSACTION of the ASAE. 29:1373-1378.

Taylor, A. W., Glotfelty, D. E., Turner, B. C., Silver, R. E., Freeman, H. P., Weiss, A. 1977. Volatilization of dieldrin and heptachlor residues from field vegetation. *Journal of Agricultural and Food Chemistry* 25(3):542-548.

Taylor, A.W. and Spencer, W.F. 1990. Volatilization and vapor transport processes. In: Cheng, H.H., Pesticide in the Soil Environment: Processes Impacts and Modeling. Number 2 in the Soil Science Society of America Book Series, SSSA, Madison, Wi. Turner, B. C., Glotfelty, D. E., Taylor, A. W. 1977. Photodieldrin formation and volatilization from grass. *Journal of Agricultural and Food Chemistry* 25(3):548-550.