BY MICHAEL P. ANDERSON

The fingerprinting of Bermudagrass DNA

Genetic relationships are key to efficient production of high-quality varieties

The fingerprinting of plant, animal and human DNA has been practiced by researchers and forensic scientist for many years, especially garnering widespread attention from notorious cases involving DNA evidence. The DNA fingerprint analysis is so powerful it's capable of distinguishing one individual from another. Each of us has a unique DNA pattern as do plant species and plant varieties.

All organisms have identifiable characteristics, which make an organism unique from all others. Physical characteristics in Bermudagrass, such as: leaf thickness or colors are obvious and readily discernable. However, some characters require detailed measurements, while others are more qualitative in nature. Some distinguishing features can be observed with little or no training, while others need close inspection by trained and experienced personnel. Many subtle differences among closely related Bermudagrasses can't be readily distinguished visually (See photo on opposite page). Another method is necessary to differentiate these Bermudas - DNA fingerprinting.

Differences among organisms are coded by their DNA. DNA is a long linear molecule consisting of a specific sequence of four distinct chemicals called nucleotides in a linear order. If the human genome were represented by letters standing for each distinct nucleotide (A, T, G, C) on a blank page, the length of the alphabetic sequence would run at least to one million pages, enough to fill 1,000 large volumes. The information in the DNA is carried in the linear sequence of the nucleotides. The DNA sequence dictates the look of an organism and how it responds to the immediate environment. This is different for every organism. Consequently, the DNA sequence can be used to distinguish one organism from another.

DNA fingerprinting is nothing more than a sophisticated technique to sample an organisms DNA sequence, projecting the differences as a kind of bar code for ready identification and comparison. Most DNA fingerprinting depends on a technique known as PCR or polymerase chain reaction. PCR was developed in the mid-1980s to efficiently amplify specific segments of DNA many, many-fold. The PCR technique uses short DNA segments composed of anywhere from six to 20 nucleotides known as primers that are complementary to segments of the target DNA. The primers figuratively scan for matches in the target DNA sequences. Once a match is found then amplification of that segment begins. If there are many matches, many segments will be amplified.

This mixture of amplified segments known as amplicons can be separated on an electrophoretic gel system which effectively sieves fragments based on size, with the largest slower moving amplicons appearing on top of the gel, and the smaller on the bottom. The gel is stained with fluorescent dyes to reveal what looks like a banding pattern or a bar code. Multiple primers can be used to scan different portions or the total genomic DNA revealing additional bar coding.

Fingerprinting with many primers is capable of differentiating even the most closely related of all organisms. Thus while two Bermudagrasses might be physically indistinguishable from each other, the DNA fingerprinting can highlight the intrinsic differences in their DNA using the PCR-based techniques.

All organisms can be finger printed and their DNA patterns stored and analyzed. Analysis of the banding pattern is performed using a variety of statistical techniques known as cluster analysis. The data is inputted in the form of presence or absence of a particular PCR amplicon or electrophoretic band and cluster analysis analyzes the data and connects those organisms that show

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Summary

- Researchers at OSU use DNA fingerprinting to evaluate the genetic background of Bermudagrass varieties from a worldwide collection.
- Understanding genetic relationships is fundamental to the efficient production of high-quality Bermudagrass varieties.
- DNA fingerprinting coupled to cluster analysis is able to distinguish and infer genetic relationships among even the most closely related organisms from each other.
- DNA fingerprinting can be used in basic and applied research, genetics, plant breeding, marker assisted selection, agricultural forensics and patenting, and ecological genetics.
similar patterns (Figure 3b). However, to be effective there must be enough similarities and differences in the pattern to reveal relationships among all tested organisms.

A number of fingerprinting techniques exist. These techniques differ in the ability to differentiate organisms, the amount of labor required, the extent of automation available, the expense of use, and nature of the specific targeted DNA segments. AFLP, DAF, SSR, RAPD are a few of the more commonly used techniques used to fingerprint DNA. All of these use PCR to amplify segments of DNA based on the DNA sequence.

In our research, we've used DAF primarily for its simplicity, low cost, ease of use and high resolution (Yerramsetty et al., 2005). Others have used more sophisticated technology to meet similar objectives (Wu et al., 2005) (Zhang et al., 1999). Sophisticated and expensive commercial packages and instrumentation exists to automate and increase the resolution of the fingerprinting procedure. Access to DNA sequencing instrumentation provides a tremendous boost in fingerprinting performance and throughput, but at a significant cost.

**HOW IT'S USED**

How has this technology been used in the past, and how might it be used in the future? We'll focus on what we and others have learned about Bermudagrasses or other species using the DNA fingerprinting techniques.
DNA fingerprinting has been used initially to look at the genetic relationship among a wide range of Bermudagrasses. Some of the first work highlighted the differences among high quality commercial cultivars and select bermudagrasses found in germplasm collections. In 1995, Caetano Anolles and other researchers surveyed 13 Bermudagrass cultivars including African, Common Bermudagrass and several interspecific hybrids for genetic relatedness using DAF. Results showed that DNA fingerprints were easily distinguishable, and the analysis showed clear genetic relationships among all bermudagrass varieties. To probe the limits of the ability to distinguish Bermudagrasses the authors fingerprinted Tifway and its irradiation induced mutant Tifway II, which presumably differed in one or a few nucleotide changes in the DNA sequence. To differentiate these very closely related varieties, the authors found it necessary to use 81 distinct primer combinations to find a one band difference among all 81 fingerprints (Caetanoanollas et al., 1995). From this early work, it was clear investigators can differentiate and draw genetic relationships even among the most closely related Bermudagrasses.

Breeders often collect from throughout the world a wide range of plant introductions in the hope of finding specific genetic traits that might be put to productive use. The genus *Cynodon* is comprised of nine species (Taliaferro, 1995). Oklahoma State University is home to a worldwide collection of Bermudagrass varieties and plant introductions that was initiated by the celebrated geneticist Jack Harlan. Charles Taliaferro and more recently Yanqi Wu, two Bermudagrass breeders at OSU, have added significantly to this collection, making it one of the most comprehensive collections of *Cynodon* germplasm in the world. In a survey of this world-wide collection using DAF fingerprinting techniques Assefa et al. 1997 (S. Assefa, 1999) examined 42 bermudas for genetic relatedness and found generally that the fingerprinting supported the taxonomic classification based on morphology by Harlan (Taliaferro, 1995). Understanding the genetic relatedness among *Cynodon* sp. and varieties gave us a better understanding the genetic make up of the *Cynodon* genus.

At times, doubts about the genetic identity of a particular cultivar surfaces. To field personnel, the cultivar doesn’t look like what it’s supposed to be. In previous work, our laboratory responded to the need to evaluate the widely used variety U3 for genetic fidelity (Anderson et al., 2001). U3 was an early success made up of Bermudas collected from golf courses in the Southern U.S. in the 1930s. U3 showed moderate cold tolerance and fine textured leaves and was a general improvement when compared to previous cultivars. Since then, U3 has been sold and marketed throughout the region.

DNA fingerprinting was employed to distinguish the current labeled U3 from presumably authentic U3 collections assembled from throughout the country. Results showed the currently labeled U3 varieties differed substantially from the presumably authentic U3 varieties (Figure 4). How these differences came about couldn’t be addressed by the fingerprinting technique, but the research underscored the need for evaluating current varieties for genetic stability and purity. Additionally, our research (unpublished) as well as others (Zhang et al., 1999) has discovered a few other discrepancies between the historical pedigree claims of several varieties and their actual genetic relationships using fingerprinting techniques.

Often times when researchers conduct experiments with particular varieties or germplasm, it’s important to understand the genetic background of the Bermudas involved. When constructing genetic mapping populations it’s essential to document the genetic background of the potential parents beforehand. The parents should differ substantially in the targeted trait while showing significant similarity in genetic background. A preliminary DNA fingerprinting survey of potential parents is the best way to do this reliably. The same can be said when selecting Bermudagrass varieties for basic research analysis. Understanding the genetic background and relationships improves experimental analysis and interpretation significantly.

**GAINING DIVERSITY**

New Bermudagrass germplasm has been and is now being collected and assembled into worldwide collections from many sources. There are areas where collections have only recently been assembled from specific geographic locations, such as Southern Asia and Southeastern Asia. Recently, Yanqi Wu brought a number of Bermudas from China adding to the OSU germplasm collection. DNA fingerprinting using AFLP technique was used to evaluate the diversity within this germplasm.

The Chinese collection seemed surprisingly diverse (Wu et al., 2006) and distinct from other Bermudas from other geographic locations around the world (Wu et al., 2004). Additional work in our laboratory easily separated the Chinese collection from all U.S. varieties tested (unpublished). Over all, the work indicated a source of significant variation in the new Chinese collection which may contain valuable genes for Bermudagrass development. Additional diversity assessments needs to be done on collections from India and other areas not surveyed previously.

The same techniques used for DNA fingerprinting such as AFLP or SSR also are used for molecular genetic analysis of specific traits. The goal here isn’t so much an analysis of diversity or genetic relatedness but for locating specific genetic elements or genes that contribute substantially to those traits. This is performed by constructing populations with significant variation in a particular trait of interest and then performing the DNA fingerprinting technique on members of the population to identify specific genetic elements that correlated with the phenotypic expression of that trait. These genetic elements are visualized as unique bands on electrophoretic gels that appear to correlate with traits of interest. The bands are valuable because they can serve as genetic markers, markers that are based on the DNA sequence rather than...
some physical characteristic of the plant.

Sophisticated computer software analysis can gauge the contribution of the DNA element associated with the marker to the genetic makeup of the phenotype. These markers can be used to increase the efficiency of selection in a process known as marker assisted selection. Marker assisted selection has been shown to be effective in enhancing germplasm improvement in a variety of cropping systems (Mackay and Powell, 2007; Tuberosa and Salvi, 2006; Yamaguchi and Blumwald, 2005). Constructions and evaluation of mapping populations and usage of molecular genetic analysis are major goals of the OSU Bermudagrass team.

Bermudagrass is an outcrossing species indicating an expected level of genetic heterogeneity within Bermudagrass populations. Typically, seeded populations consist of a range of individuals that differ genetically. The genetic diversity within the population may be wide or narrow depending on the way the population was constructed originally. A wide genetic base consists of many individuals that differ substantially from each other. When we characterize genetic populations we must evaluate the entire population, sampling a representative number of individuals. So far, this has rarely if ever been performed on seeded Bermudagrasses.

DNA fingerprinting of individuals within a population provides information concerning the genetic make-up of that population. The individual makeup of the population might change with time depending on natural selection and genetic inflow from neighboring Bermudas. To observe these shifts, DNA fingerprinting can be used to document and track alterations in population make-up of seeded Bermudagrasses under a variety of environmental conditions throughout time. So far, little is known concerning this aspect of Bermudagrass culture, which needs more investigation, especially considering the emergence and use of fertile seeded populations in the Bermudagrass industry.

AGRICULTURAL FORENSICS AND PATENTING

DNA fingerprinting also can be used in areas of agricultural forensics. One case illustrates this use. Years ago, a farmer was concerned about theft of Bermudagrass hay bales from his farm. The farmer had several suspected culprits in mind and contacted us to determine if DNA could be used to support a claim prior to legal action. To prove the claim, samples would have

![Figure 3a. Cluster analysis](https://example.com/figure3a)

**Figure 3a. Cluster analysis**

**MHP analysis**

- A12205
- A12250
- A19198
- Pat
- OKC41-8
- OKC70-18
- Tifton10
- Baby
- Tifgreen
- Midlawn
- Quickstand
- Tifport
- Tifway

![Figure 3b. Cluster analysis](https://example.com/figure3b)

**Figure 3b. Cluster analysis**

- Arizona Common
- CD90160
- Jackpot
- Majestic
- Savannah
- Southern Star
- Sundevil
- Mohawk
- Riveria
- Mirage
- Sydney
- Pyramid
- Numex Sahara
- Transcontinental
- Princess
- SWI-11
- Yukon

Further supporting evidence including cultural histories and practices among the implicated parties would have to be provided – a significant and costly undertaking. The evidence would have to be evaluated by an expert using quantitative and statistical models before a legal opinion could be constructed. In this case, the effort appeared too costly in terms of time and money; however, there might be cases where the expense and effort is justifiable.

Finally, DNA fingerprinting can have an
impact in the area of patent protection. Many years and efforts are expended to develop commercial varieties. Institution have a substantial investment in terms of developmental cost, and are increasingly desirous of recovering some of that cost through plant variety protection, and the collection of royalties from consumers. To support the patent application process, differences in morphology, cultural characteristics and pedigree needs to be presented to distinguish the proposed variety from those that are currently available. DNA fingerprinting is currently being used on a limited basis to document the genetic differences of new varieties in the patent process. Any infringement on the patent would have to use the DNA fingerprints and other characteristics to justify a patent infringement lawsuit. The process may be costly and subject to interpretation by experts, but may be worth the effort when the stakes are large.

**ECOLOGICAL GENETICS**

Ecological studies in the natural environment are often times helpful in distinguishing among ecotypes that might differ in desirable or undesirable characteristics. At OSU, we collaborated with a project seeking to identify various ecotypes of *Sericea lespedeza*, a major introduced invasive species that threatens forage production on natural pasture lands in Oklahoma (Farris, 2004). The idea was to look at genetic background of the different ecotypes and its relationship to the ability to control this problem pest. Understanding the genetic base of the *Sericea lespedeza* populations might be an important element in designing more effective control methods.

DNA fingerprinting is a valuable technology that’s being used to assist producers, breeders, geneticist and researchers evaluate Bermudagrass populations and germplasm for genetic diversity and background. Information from DNA fingerprinting techniques allow researchers to make informed decisions concerning progress in developing high quality bermudagrass lines. DNA fingerprinting technology remains a powerful technique in assessing the genetic diversity of Bermudagrasses worldwide and at protecting plant varieties from infringement. Our projects have been involved in using DNA fingerprinting to improve Bermudagrass more. GCI

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The author acknowledges the contribution of Charles Taliaferro, Ph.D., for providing access to Bermudagrass collections and for discussions and insight related to Bermudagrass genetics and breeding; Praveen Yerramsetty and Carole Anderson for their technical skills developing the data on which much of the work depends. He appreciates the USGA and the Oklahoma State Agricultural Experiment Station for providing funds for the execution and completion of this and other works associated with Bermudagrass improvement.

Editor’s note: References for this article can be found at www.golfcourseindustry.com.

**IMPACT ON THE BUSINESS**

**Fingerprinting takes the guesswork out of turf identification**

From a crop perspective, genetics plays an important role in purity of product, yield and how inputs such as pesticides and fertilizers will react.

DNA fingerprinting in turfgrass allows superintendents and breeders the ability to distinguish between plant varieties, particularly if a choice needs to be made between two or more varieties.

“Fingerprinting looks at the DNA of a particular variety,” says Michael Anderson, Ph.D. “It doesn’t really check or tell the health of plant, but can distinguish one from the other.”

While many superintendents look at variations of color to determine the purity of a turfgrass plant, the only way to determine whether it’s a pure variety is through fingerprinting, which can determine this without question.

This technology, introduced in the 1980s, aids in the identification rather than the maintenance of turfgrass, although fingerprinting is helpful in determining whether or not a turfgrass variety has been contaminated.

Anderson relates a situation in which superintendents and others were wondering whether or not a variety was really what they said and thought it was. “We did a bunch of tests on a variety that we thought was U3 Bermudagrass. That’s how it was labeled, but it didn’t look like the original U3. To nail it down, we conducted fingerprinting and found it was absolutely different from U3.”

Fingerprinting answers academic questions such as which variety is better adapted for local conditions. Rather than trusting the label or counting on visual gut checks, fingerprinting is the only method that will provide superintendents with an absolute.

While superintendents can make do without fingerprinting, breeders, sod producers and researchers rely on the technology to ascertain what variety they’re working with. “In turf plots, I want to know if a variety has been overgrown by a contaminant,” Anderson says. “Contamination can impact the production of new varieties.”

Anderson says he’s been using these techniques for the past 10 years in his Bermudagrass research because even he can’t always distinguish between varieties. He can’t leave it to chance when beginning a research project. He needs to know the genetic background of what he’s working with.

Breeders also will use fingerprinting to compare the genetics of Bermudagrass collected worldwide. Superintendents most often can test Bermudagrass on their courses visually, but if contamination is an issue, fingerprinting will help you get a straight answer. GCI
Improving turf quality
How Bermudagrass genotypes respond to mowing height and nitrogen or growth regulators

TifSport, a high turf quality and fine-textured interspecific triploid (2n=3x=27 chromosomes) Bermudagrass hybrid, was released in 1995 (Hanna, Carrow and Powell, 1997). It's genetic purity, improved cold resistance, superior sod strength, pest resistance, turf density and improved traffic tolerance have made it a popular choice to plant on golf courses, athletic fields, lawns and landscape areas.

L. Cella and other researchers (2005) found that golf ball lie varied among Kentucky bluegrass cultivars and the number of plant tillers showed the highest correlation to ball lie. It was brought to our attention that, although TifSport performed well on golf courses, high handicap golfers wanted to see the ball with a higher lie. Therefore, we initiated this study to see if nitrogen levels combined with growth regulators would increase the lie at four different mowing heights or schedules. We used a modification of an instrument (see top-left photo on page 96) described by Cella and others (2004) to measure ball lie.

EXPERIMENTAL PROCEDURES
TifSport Bermudagrass (plot established in 2004) and two experimental vegetatively propagated Bermudagrasses – Tifton 11 and Tift No. 4 (ST-5) – were established in 2005. Tifton 11 and Tift No. 4 also were selected for testing because both of these experimental cultivars show potential for golf course use. The design was a strip plot test with four replications. Treatments included three nitrogen levels combined with Primo (trinexapac-ethyl) and Cutless (flurprimidol) and four mowing heights. A treatment with one pound of nitrogen per 1,000 square feet per month plus Primo was considered a general practice used by golf course superintendents.

The nitrogen/Primo/Cutless treatments were:
• 0.5 pound of nitrogen per month per 1,000 square feet
• 1 pound of nitrogen per month per 1,000 square feet
• 1.5 pounds of nitrogen per month per 1,000 square feet
• 1 pound of nitrogen per month per 1,000 square feet plus Primo
• 1.5 pounds of nitrogen per month per 1,000 square feet plus Primo
• 1 pound of nitrogen per month per 1,000 square feet plus Primo plus Cutless
• 1.5 pounds of nitrogen per month per 1,000 square feet plus Primo plus Cutless

Primo was applied at nine ounces per acre in Primo-only treatments and at four ounces per acre in Primo/Cutless treatments. Cutless was applied at four ounces per acre. Treatments were applied once a month during the growing season, May through October.

Mowing heights were:
• 0.5 inch (12.5 mm), twice a week
• 1 inch (25 mm), twice a week
• 1.5 inches (37.5 mm), twice a week
• 1.5 inches (37.5 mm), once a week

The mowing heights were selected to approximate practices used in various areas of the golf course. Quality and color ratings usually were taken at the end of the month before the new treatments were applied.

BALL LIE
Ball lie measurements were taken by dropping two golf balls into each plot from a height of six feet and then measuring the distance the ball sunk into the turf (see top-right photo on page 96). Data on turf quality were collected in 2005 and 2006. Data on ball lie were collected in 2005 (three dates) and 2006 (three dates) for TifSport but only in 2006 (one date) for Tifton 11 and Tift No. 4 (ST-5). Rating used ranged from one to nine with nine being the best turf quality. A rating of at least seven is needed for acceptable turf quality.

A golf ball is 1.65 inches in diameter. The values listed in tables for ball height indicate the number of millimeters the ball sank into the surface of the grass. Therefore, the smaller the number, the higher the ball lie. All ratings and ball-lie measurements were rounded to the nearest whole number.

Summary points

- Treatments with one or 1.5 pounds of nitrogen per 1,000 square feet produced similar turf quality and color in TifSport, Tifton 11 and Tift No. 4.
- Treatments with nitrogen plus Primo or nitrogen plus Primo and Cutless didn't have considerable effects on improving turf quality or color.
- An application of Primo or Primo plus Cutless produced a denser turf that provided a higher ball lie in TifSport. Ball lie in Tifton 11 and Tift No. 4 were similar for all treatments.
whole number because decimal values have little practical value. An analysis of variance was used to determine the effects of various treatments on turf quality and ball lie. Fisher's LSD test was used to determine differences between treatments (SAS Institute, Cary, N.C.).

TURF QUALITY
There were only small differences in overall turf quality except for the 0.5-pound-of-nitrogen-per-1,000-square-feet treatment in which turf quality was reduced for TifSport and for Tifton 11 in 2005. We also observed lighter green color (data not shown) for the 0.5 nitrogen treatment for TifSport and Tifton 11, but not for Tift. No. 4. We observed a little discoloration in the Cutless treatments for a few days after treatment. Cutless appeared to discolor Tift 97-4 more than the other genotypes, probably because this cultivar is the most naturally dense grass of the three tested. We observed the least discoloration in Tifton 11, and it's the most coarse grass of the three tested.

One pound of nitrogen per 1,000 square feet per month appeared adequate for maintaining desirable turf quality in all three grasses (see comparison photo, bottom right, on page 96). However, 0.5 pound of nitrogen per 1,000 square feet per month might be adequate for Tift No. 4, a dense, naturally dark green, shade-resistant genotype. Neither Primo nor Cutless improved overall turf quality in this test. However, clipping removal (not measured in this test) probably would have been reduced by the growth regulators.

Turf quality tended to improve for TifSport from 2005 to 2006 as the turf 'matured'. Treatments with Cutless (at the rate used) caused browning and swirling of the turf at 0.98 inch and 1.46 inches mowing heights for about a week after treatment in TifSport and Tift 97-4, which was especially pronounced at the October treatment. There were only small differences in turf quality because of mowing heights (see table 2 at right).

Table 1. Mean turf quality ratings for TifSport, Tifton 11 and Tift No. 4 in 2005 and 2006.

<table>
<thead>
<tr>
<th>Treatment</th>
<th>TifSport</th>
<th>Tifton 11</th>
<th>Tift No. 4</th>
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<tbody>
<tr>
<td>0.5 N</td>
<td>7</td>
<td>7</td>
<td>7</td>
</tr>
<tr>
<td>1.0 N</td>
<td>7</td>
<td>8</td>
<td>8</td>
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<tr>
<td>1.5 N</td>
<td>7</td>
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<td>8</td>
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<tr>
<td>1.0 N + P</td>
<td>7</td>
<td>8</td>
<td>8</td>
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<tr>
<td>1.5 N + P</td>
<td>7</td>
<td>8</td>
<td>8</td>
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<tr>
<td>1.0 N + P + C</td>
<td>7</td>
<td>7</td>
<td>8</td>
</tr>
<tr>
<td>1.5 N + P + C</td>
<td>7</td>
<td>8</td>
<td>8</td>
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<tr>
<td>LSD - 5%</td>
<td>1</td>
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<td>1</td>
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</tbody>
</table>

Turf quality: 9=best, 7=acceptable quality
N=Nitrogen, P=Primo and C=Cutless

Table 2. Mean turf quality ratings for TifSport, Tifton 11 and Tift No. 4 in 2005 and 2006.

<table>
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<tr>
<td>0.5 - 2x/wk</td>
<td>7</td>
<td>8</td>
<td>8</td>
<td>8</td>
<td>7</td>
<td>7</td>
</tr>
<tr>
<td>1.0 - 2x/wk</td>
<td>7</td>
<td>8</td>
<td>7</td>
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<td>7</td>
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<td>1.5 - 2x/wk</td>
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<td>1.5 - 1x/wk</td>
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<td>7</td>
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<tr>
<td>LSD - 5%</td>
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<td>1</td>
</tr>
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</table>
**BALL HEIGHT**

The nitrogen level had little effect on keeping the golf ball from sinking into the grass (see table 3 on page 97). All combinations of nitrogen, Primo and Cutless were effective in improving ball lie in TifSport. As TifSport (planted in 2004) matured from 2005 to 2006, the ball lie improved. Treatments had almost no effect on ball lie in Tifton 11 and Tift No. 4. Tifton 11 is quite vigorous - producing dense turf - so it is little affected by nitrogen inputs.

Top left: Researchers used a modification of an instrument described by L. Cella and other researchers. Top right: Ball lie measurements were taken by dropping two golf balls into each plot from a height of six feet and then measuring the distance the ball sank into the turf. Bottom left: As TifSport (planted in 2004) matured from 2005 to 2006, the ball lie improved (right). Treatments had almost no effect on ball lie in Tifton 11 (left). Bottom right: One pound of nitrogen per 1,000 square feet per month appeared adequate for maintaining desirable turf quality in all three grasses.

*Photos: Wayne Hanna*

**IMPACT ON THE BUSINESS**

*They know what they like...*

Golfers - even high-handicappers - are a notoriously picky breed. When it comes to turf conditions, the old statement about art appreciation holds true for even an average hacker: They don't know much, but they know what they like.

One of the things they appear to like is a lie in the rough where the ball sits up, making it easier to make contact and rescue themselves from lousy shots. In short, even though they've hit the ball where they're not supposed to, they believe lies where the ball sits down are bad.

Thus, Wayne Hanna, Ph.D., and his team at the University of Georgia - the home of the various Tif species - took a look at how the ball rests when dropped on their turf. More specifically, the question is whether nitrogen and plant growth regulator inputs impacted the way the ball sits up on TifSport.

The bottom line of the study - which was largely funded by the USGA - is that growth regulators have a positive impact on how high a ball will sit in TifSport mowed at between 0.5 inch and 1.5 inches.

**Trend**

More superintendents are using PGRs in the rough than ever before, according to chemical company representatives. The primary value is reduced growth, which translates to less mowing, thus lowering labor costs. But a secondary benefit such as improved lies provides a nice opportunity to improve the playability of the course for mediocre players who despise bad lies.

**Cost/benefit**

PGRs aren't inexpensive, but the documented benefits continue to multiply as turf researchers and superintendents experiment with them. Regular treatments using the Primo/Cutlass combination described would add more than $5,000 annually to a facility's PGR budget.

However, this is offset by the potential for:

- Reduced mowing costs;
- Thicker turf;
- Upright growth (better lies);
- Better annual bluegrass performance (seedheads); and
- Fewer clippings.

**Bottom line**

Southeastern courses catering to mid- to high-handicap golfers could consider a program like this to manage TifSport fairways and roughs, thus improving ease of play and perhaps speeding up play to accommodate more rounds and make those picky golfers happy. GCI
apparently can mature turf soon after planting. Tift No. 4 is a naturally dense turf. The highest ball lie was achieved with 1.5 pounds of nitrogen combined with Primo and Cutless.

It appears from these results that one pound of nitrogen plus Primo can produce a good ball lie. Users would need to decide for themselves whether the slight improvements in ball lie are worth the extra cost of another half pound of nitrogen and/or Cutless per month. A lower level of Cutlass also might prevent some of the discoloration observed in this study.

Mowing at one-half inch twice a week produced the best ball lie in all three Bermudagrasses (see table 4 at right). The lowest mowing height produced the most dense turf. As mowing height increased and mowing frequency decreased, the ball sank further into the grass for TifSport and Tifton 11, and for Tift No. 4 going from the one-half inch to one inch mowing height. There were no differences in ball lie at the 1.5 inch mowing heights for Tifton 11 and Tift No. 4. The ball lie in TifSport improved from 2005 to 2006, probably because of the production of a more mature turf.

Another consideration in this mowing height is how far the bottom of the ball is from the ground for the various mowing heights (numbers in parenthesis in table 4). Although the ball sinks less into the grass at the half-inch mowing height, the ball is further from the ground at the one inch and 1.5 inches mowing heights.

Treatments with Primo or Primo plus Cutless were the most effective for preventing the golf ball from sinking into TifSport. The nitrogen level by itself appeared to have little effect on ball lie. Tifton 11 was exceptional at all treatment levels and mowing heights for keeping the ball from sinking into the turf.

Wayne W. Hanna, Ph.D., is a professor in the department of crop and soil sciences at the University of Georgia, Tifton Campus. He can be reached at 229-386-3184.

The author wishes to thank the USGA for financial support to conduct the research and to Raymond Cooper for requesting Cutless. Appreciation is expressed to Patrick O’Brien, director of the southeast region USGA Green Section and to Jimmy Allen from Pike Creek Turf for discussions regarding need for the research, and to Larry Baldree and Amanda Webb for technical assistance.

Table 3. Mean ball height measurements (mm) for TifSport, Tifton 11 and Tift No.4 in 2005 and 2006.

<table>
<thead>
<tr>
<th>Ball height- mm*</th>
<th>TifSport</th>
<th>Tifton 11</th>
<th>Tift No._4</th>
</tr>
</thead>
<tbody>
<tr>
<td>Treatment</td>
<td>2005</td>
<td>2006</td>
<td>2006</td>
</tr>
<tr>
<td>0.5 N</td>
<td>24</td>
<td>14</td>
<td>5</td>
</tr>
<tr>
<td>1.0 N</td>
<td>21</td>
<td>14</td>
<td>5</td>
</tr>
<tr>
<td>1.5 N</td>
<td>20</td>
<td>13</td>
<td>5</td>
</tr>
<tr>
<td>1.0 N + P</td>
<td>14</td>
<td>10</td>
<td>5</td>
</tr>
<tr>
<td>1.5 N + P</td>
<td>16</td>
<td>9</td>
<td>5</td>
</tr>
<tr>
<td>1.0 N + P + C</td>
<td>15</td>
<td>10</td>
<td>5</td>
</tr>
<tr>
<td>1.5 N + P + C</td>
<td>11</td>
<td>8</td>
<td>5</td>
</tr>
<tr>
<td>LSD-5%</td>
<td>3</td>
<td>2</td>
<td>1</td>
</tr>
</tbody>
</table>

N=Nitrogen, P=Primo, and C=Cutless

* The smaller the number, the higher the ball lie.

Table 4. Mean ball height measurements (mm) for TifSport, Tifton 11 and Tift No. 4 in 2005 and 2006.

<table>
<thead>
<tr>
<th>Ball Height-mm†</th>
<th>TifSport</th>
<th>Tifton 11</th>
<th>Tift No. 4</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mowing height</td>
<td>2005</td>
<td>2006</td>
<td>2006</td>
</tr>
<tr>
<td>0.5 - 2x/wk</td>
<td>8 (4)</td>
<td>5 (7)</td>
<td>3 (9)</td>
</tr>
<tr>
<td>1.0 - 2x/wk</td>
<td>10 (15)</td>
<td>8 (17)</td>
<td>5 (20)</td>
</tr>
<tr>
<td>1.5 - 2x/wk</td>
<td>24 (13)</td>
<td>13 (24)</td>
<td>6 (31)</td>
</tr>
<tr>
<td>1.5 - 1x/wk</td>
<td>28 (9)</td>
<td>17 (20)</td>
<td>6 (31)</td>
</tr>
<tr>
<td>LSD - 5%</td>
<td>2</td>
<td>2</td>
<td>1</td>
</tr>
</tbody>
</table>

† Distance (mm) the ball sank into the grass.
‡ Distance (mm) from the ground to the bottom of the golf ball.

Literature cited